

stop-PRISM integration allele in the *rb1* tumor suppressor recapitulates developmental and cellular neural progenitor loss of function phenotypes

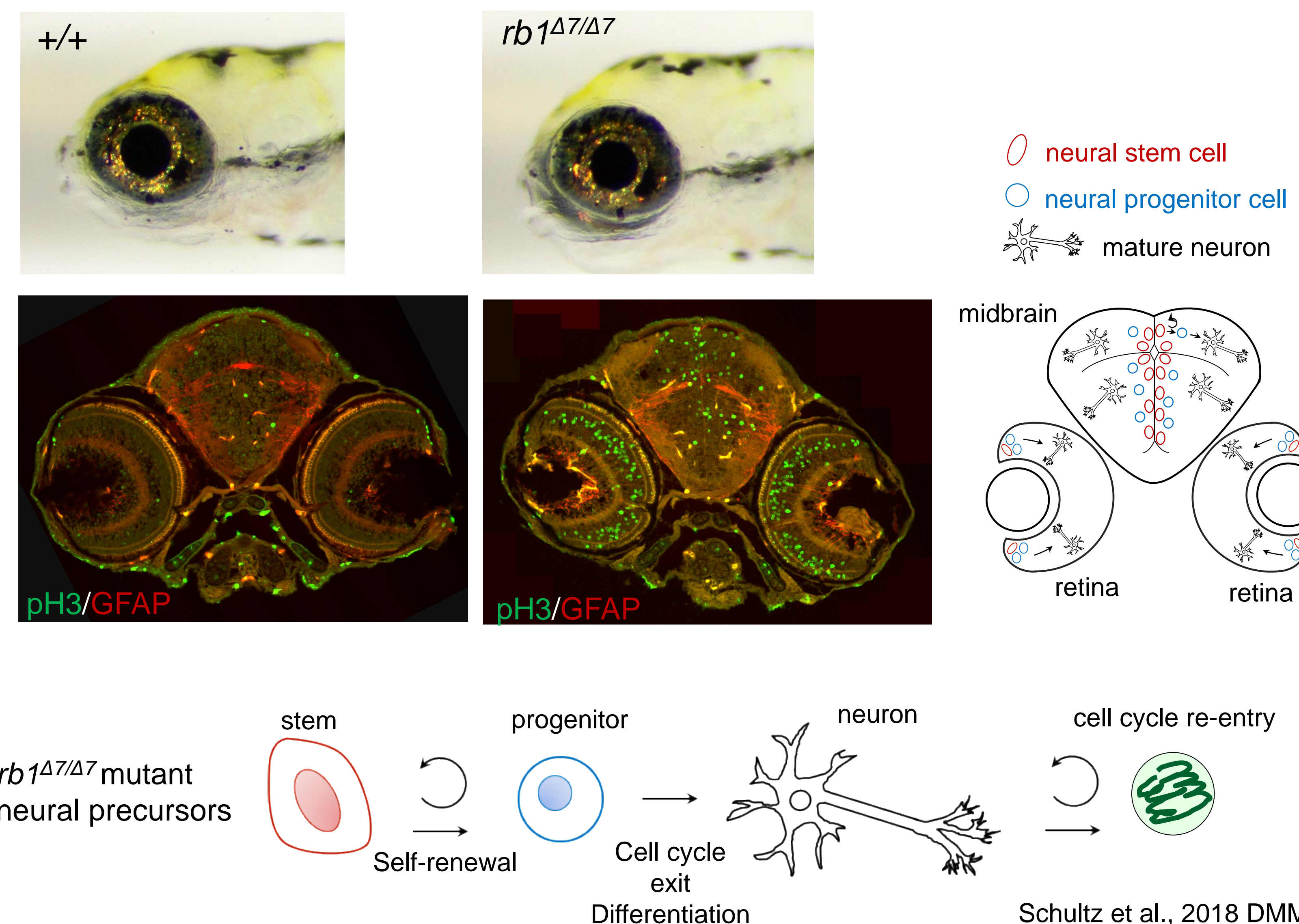
Austin Winker^{1,2}, Wes Wierson¹, Sekhar Kambakam^{1,4}, Steve Ekker^{3,4}, Karl Clark^{3,4}, Jeff Essner^{1,4}, Maura McGrail^{1,4}

¹Genetics, Development and Cell Biology, Iowa State University; ²Undergraduate Major in Genetics, Iowa State University; ³Mayo Clinic; ⁴Iowa State/Mayo Clinic Alliance for Genome Engineering

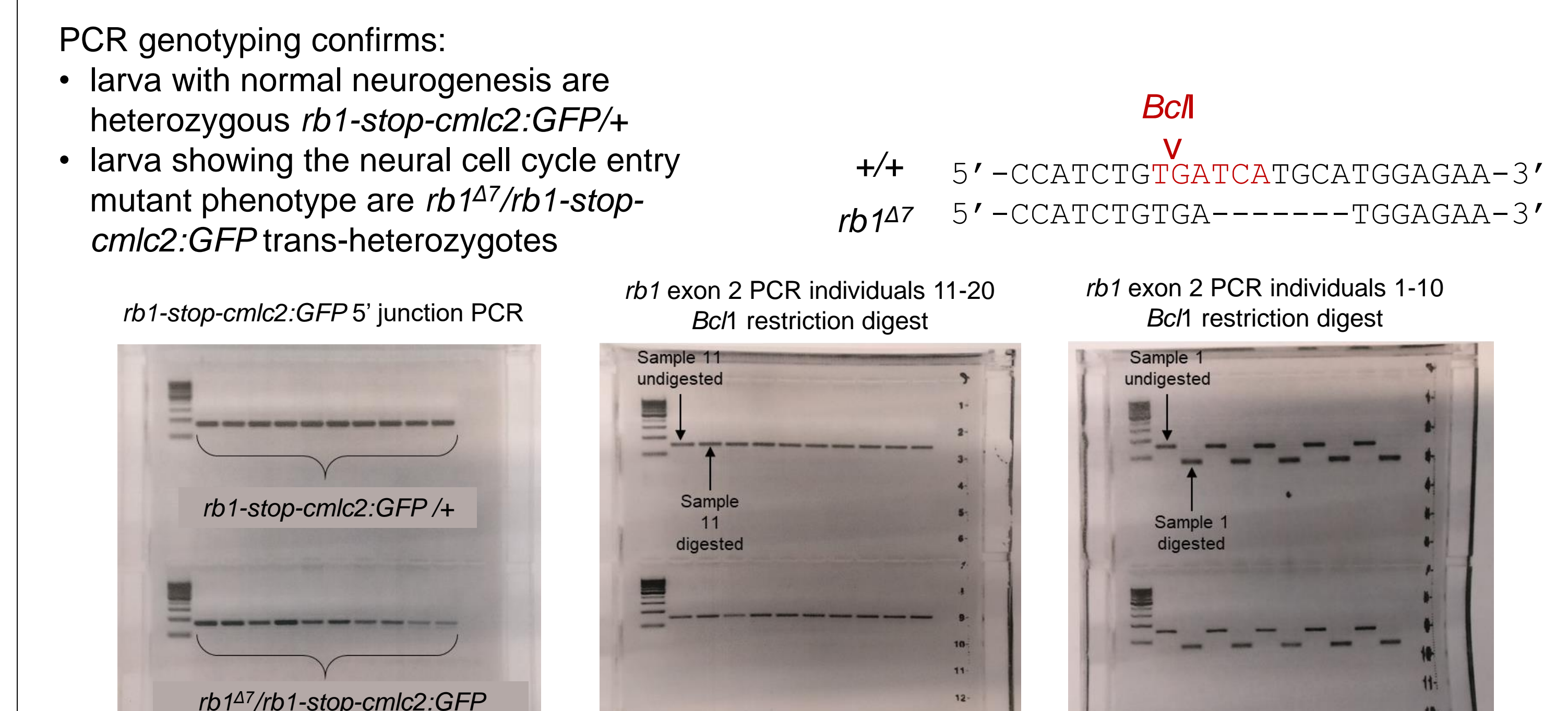
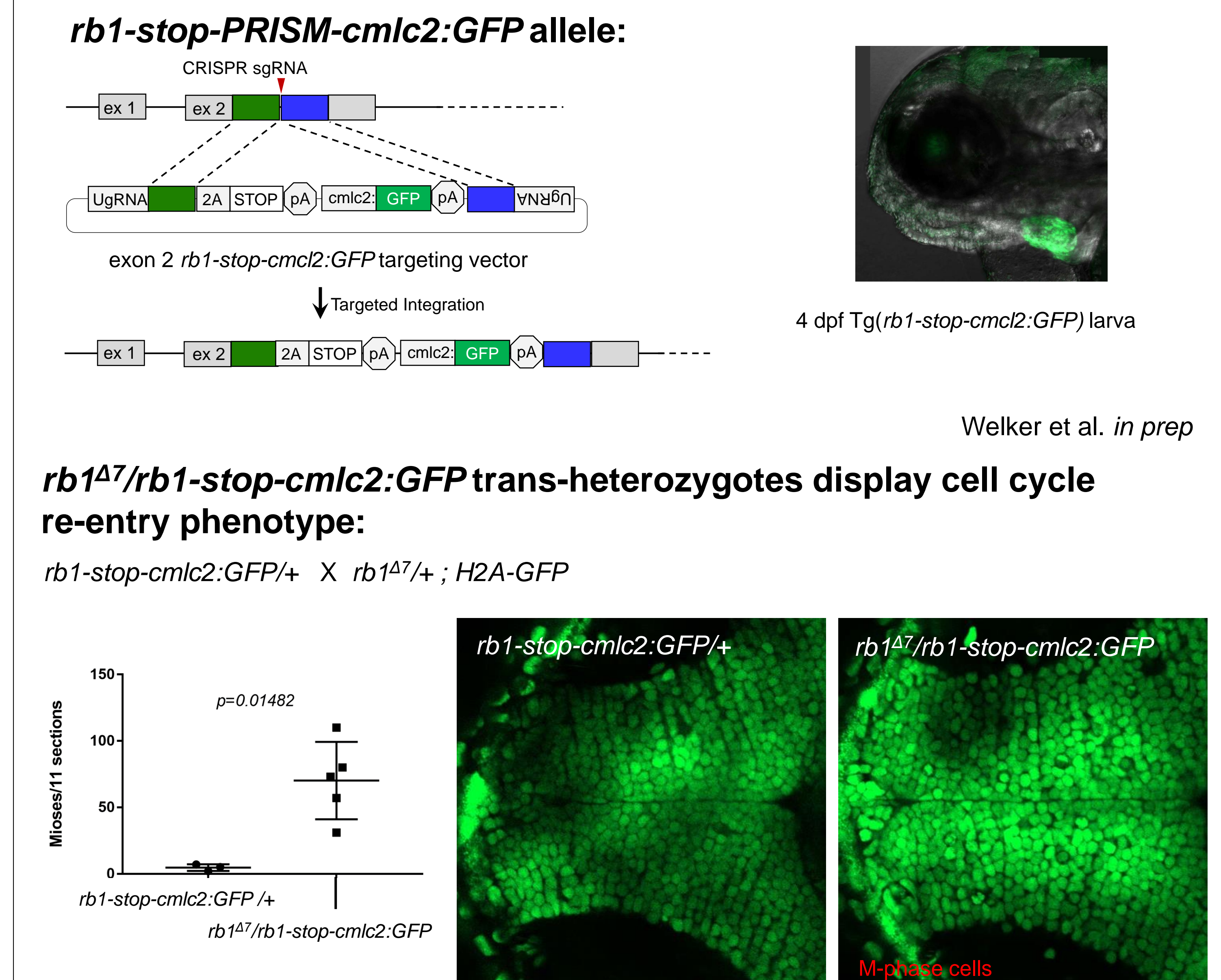
Abstract

To simplify screening and genotyping of mutant embryos we generated a set of vectors for CRISPR/Cas9 targeted integration called pPRISM, which contain secondary markers that allow for simple visual genotyping (plasmids for Precise Integration with Secundary Markers). The plasmids are designed for integration using short homology arms as we reported previously (Wierson et al., 2019 bioRxiv). The stop-PRISM vectors have a cassette for transcriptional termination followed by a secondary marker for heart, lens or hatching gland-specific fluorescent reporter expression. Targeted integration into an early exon is predicted to cause early transcription termination and create a loss of function mutation. We isolated an *rb1-stop-PRISM-cmlc2:GFP* line and tested whether the integration allele behaved as a loss of function mutation in trans with our previously published 7 bp indel allele *rb1 Δ 7is54*. The gross morphology of *rb1-stop-PRISM-cmlc2:GFP/rb1 Δ 7is54* trans heterozygotes is relatively normal, however the larvae fail to develop a swim bladder and are lethal, similar to *rb1 Δ 7is54/ Δ 7is54* homozygotes. To examine if the *rb1-stop-PRISM-cmlc2:GFP* allele disrupts *rb1* function at the cellular level we used live embryo confocal imaging of H2A.F/Z:GFP chromatin in the developing optic tectum to follow neural progenitor cell cycle dynamics. At 5 dpf, *rb1-stop-PRISM-cmlc2:GFP/rb1 Δ 7is54* mutant neural progenitors re-enter the cell cycle and condense their chromatin. The cells then appear to arrest in prophase, and fail to align the chromosomes at the metaphase plate and progress through the cell cycle, as we previously found for *rb1 Δ 7is54/ Δ 7is54* homozygotes (Schultz et al., 2018). PCR genotyping of the imaged embryos revealed the expected genotypes matched the observed phenotypes. Quantification of the neural progenitor mutant phenotype is in progress. Future analysis will compare phenotype and quantification data between our *rb1-stop-PRISM-cmlc2:GFP/rb1 Δ 7is54* trans heterozygotes and homozygous *rb1-stop-PRISM-cmlc2:GFP/stop-PRISM-cmlc2:GFP* embryos. Our results confirm integration of the pPRISM cassette can create loss of function mutations that recapitulate the phenotype of standard indels in the tumor suppressor *rb1*, both at the morphological and cellular level. stop-PRISM loss of function alleles will be useful genetic tools for rapid genotyping in combination with more complex edited alleles to study tissue or cell type specific gene function.

2. RB is required for neural progenitor cell cycle exit and differentiation during zebrafish neural development



4. *rb1* exon2 –stop-PRISM-cmlc2:GFP allele generated by CRISPR short homology directed targeted integration recapitulates *rb1* mutant neural progenitor cell cycle defects



Summary

- Knock-in stop-PRISM *rb1-stop-cmlc2:GFP* allele generated by precision targeted integration 1. allows for simple visual genotyping and 2. produces loss of functions phenotypes consistent with *rb1* indel mutation.
- Visual genotyping plus live imaging allows phenotyping of *rb1* mutant neural cellular phenotypes – two color alleles offer potential for genotyping without PCR.
- Work is ongoing to determine the interaction of *rb1* and *tp53* in neural progenitor proliferation and differentiation, and the pathways blocking progression and differentiation of *rb1* mutant neural progenitors.