

RNA-binding protein Alan shepard regulates fat storage homeostasis

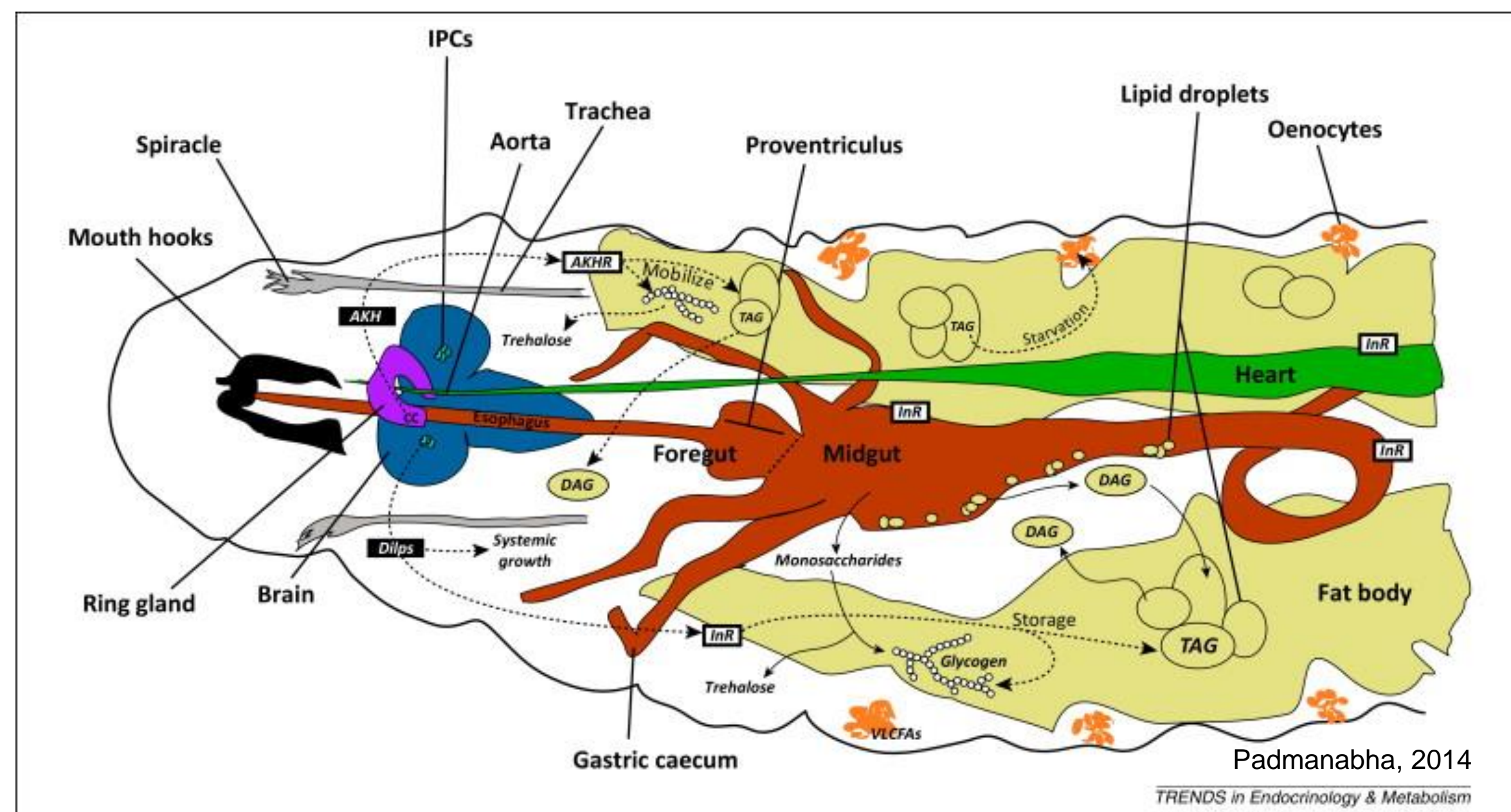
Claire Gillette, Kelsey Hazegh, Tânia Reis

University of Colorado Anschutz Medical Campus | Department of Medicine | Division of Endocrinology, Metabolism, and Diabetes | Molecular Biology Program | Aurora, CO, USA



Abstract

Metabolism is an integrated, multi-organ process, and is best studied within the context of the whole organism. Mounting evidence points to an important yet poorly understood role for genetic background in the control of organismal fat levels. The RNA-binding protein Alan shepard (*shep*) has no characterized role in metabolism. We used tissue-specific RNAi to determine in which organs *shep* is required for regulation of organismal fat. We find that knockdown of *shep* in the brain phenocopies the high-fat phenotype of the mutant and drives changes in the complex metabolic behaviors of feeding and activity. Additionally, knockdown of *shep* in the fat body results in a lean phenotype accompanied with a modest increase in activity. *Shep*'s genomic locus codes for 8 mRNA and 6 protein isoforms. Thus far, we have shown that fat body-specific overexpression of the *Shep*-RE isoform increases organismal fat levels, suggesting a positive associate between *Shep*-RE expression and the fat body's propensity to store fat. We show that the RNA Recognition Motifs are necessary for the metabolic function of *Shep*-RE. We further find that *Shep* transcript and protein levels are regulated in a nutrient-dependent manner. As the nutritional content of the diet increases, *shep* mRNA and protein expression decreases in the fat body. This suggests that *shep* is regulated by a nutrient sensing pathway. Our work is now focused on determining the role of *shep* isoforms in different organs in regulating overall organismal energy metabolism, focusing here on *Shep* function in the fat body.



Drosophila as a model for metabolic disease

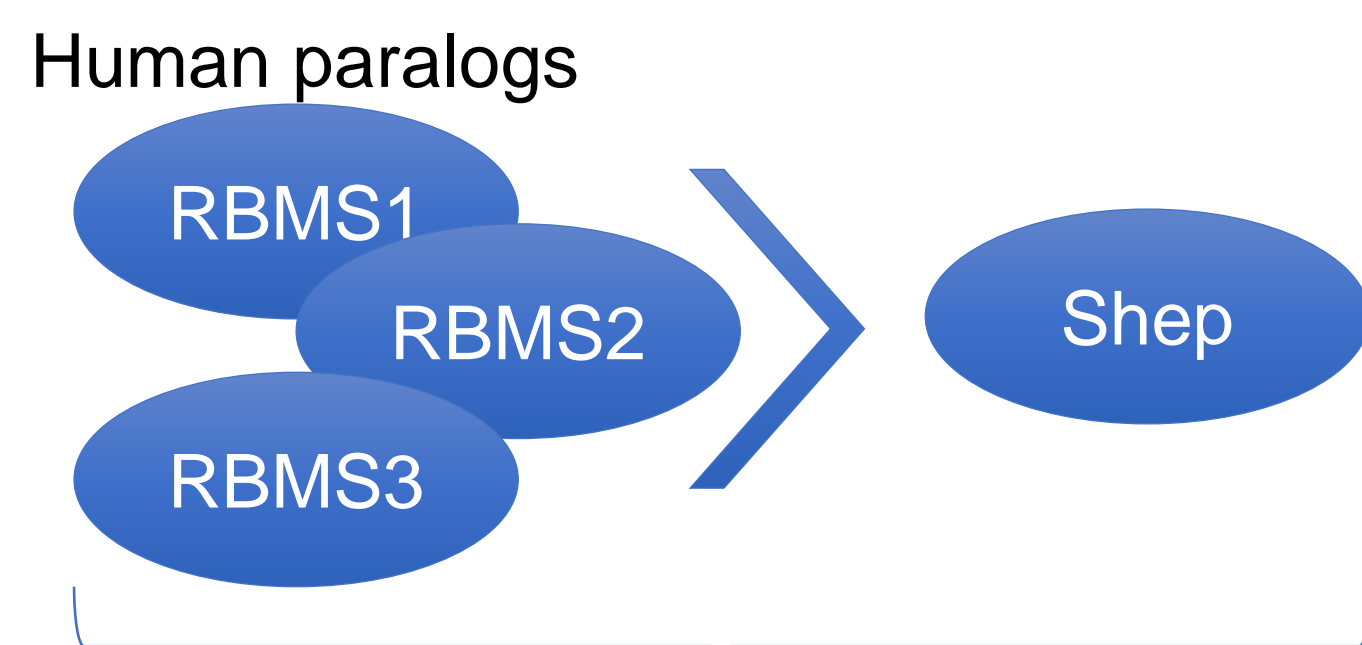
Basic metabolic pathways are highly conserved; glycolysis is very similar from yeast to humans. *Drosophila* are an advantageous model because they share many metabolic organs with mammals—brain, gut, and fat body (FB; glycogen and TGs storage organ). They also share many complex inter-tissue signaling pathways such as the gut-brain axis and neuropeptide/signal peptide signaling between metabolic organs. For example, *Drosophila* have Insulin-like peptides (dILPs) and AKH (glucagon homologue) to regulate circulating sugars.

Forward genetic screen identified RNA-binding protein Alan shepard (Shep) as obesity predisposing gene

Mutant gene	Buoyancy score	Molecular function
<i>shep</i>	+++	mRNA binding

Reis, 2010

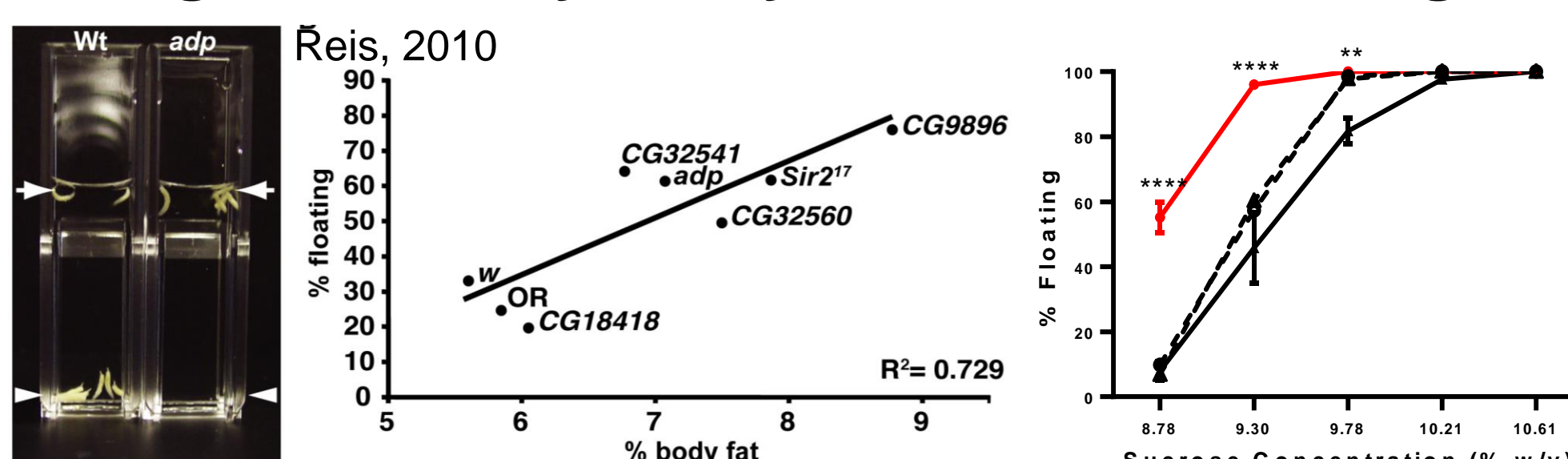
Human ortholog of Shep (RBMS) is implicated in Type 2 Diabetes Risk



Orthologs

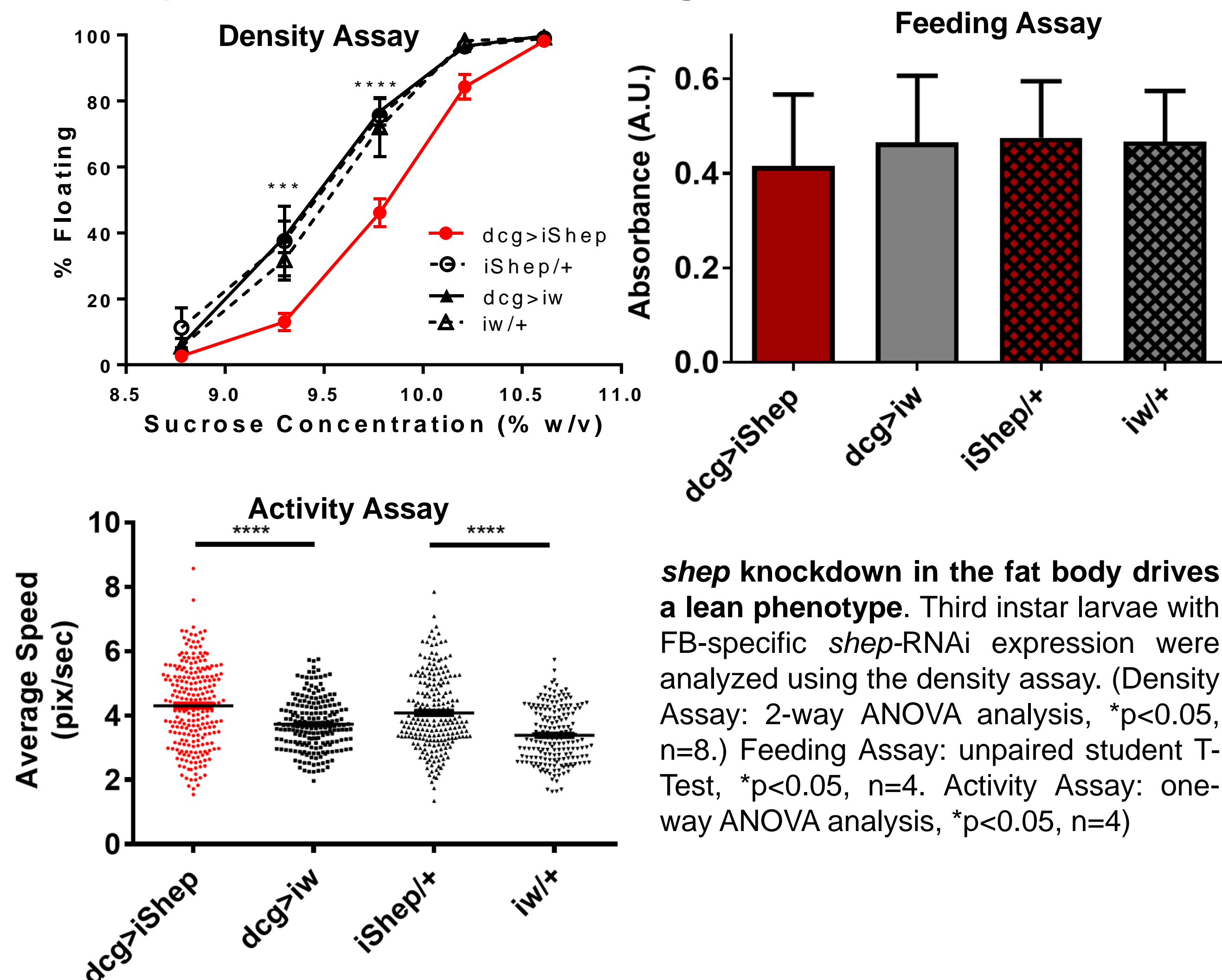
GWAS identified RBMS1 rs7593730 and rs6718526 as significantly associated with T2D risk (Campbell, 2012 and Kazakova, 2018).

Using the density assay to assess fat storage



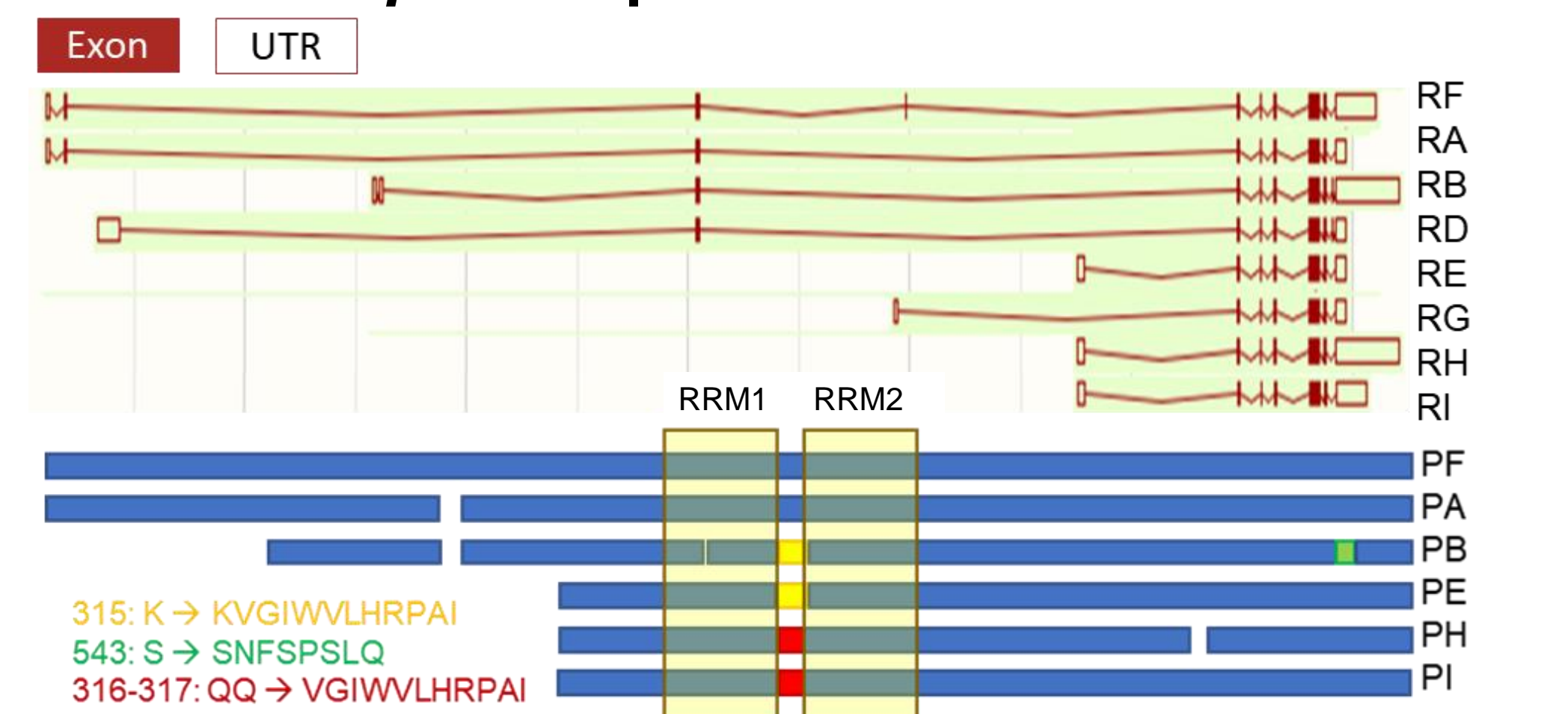
Fat tissue is less dense than lean tissue. As % body fat increases, the % larvae floating increases. In the last graph the red-line genotype is comparatively more fat than the black-line genotypes and controls.

FB-specific *shep*-RNAi drives a lean phenotype that is independent of behavioral changes



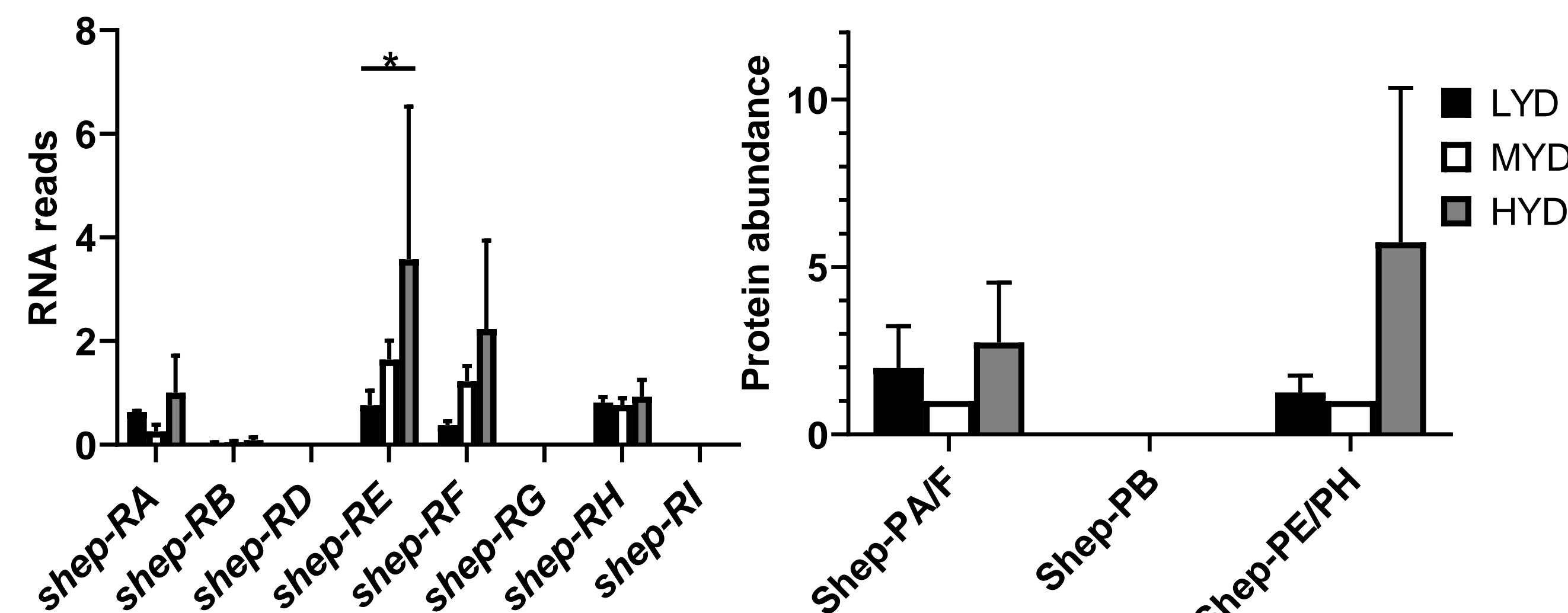
***shep* knockdown in the fat body drives a lean phenotype.** Third instar larvae with FB-specific *shep*-RNAi expression were analyzed using the density assay. (Density Assay: 2-way ANOVA analysis, *p<0.05, n=8.) Feeding Assay: unpaired student T-Test, *p<0.05, n=4. Activity Assay: one-way ANOVA analysis, *p<0.05, n=4)

The *shep* locus produces several isoforms



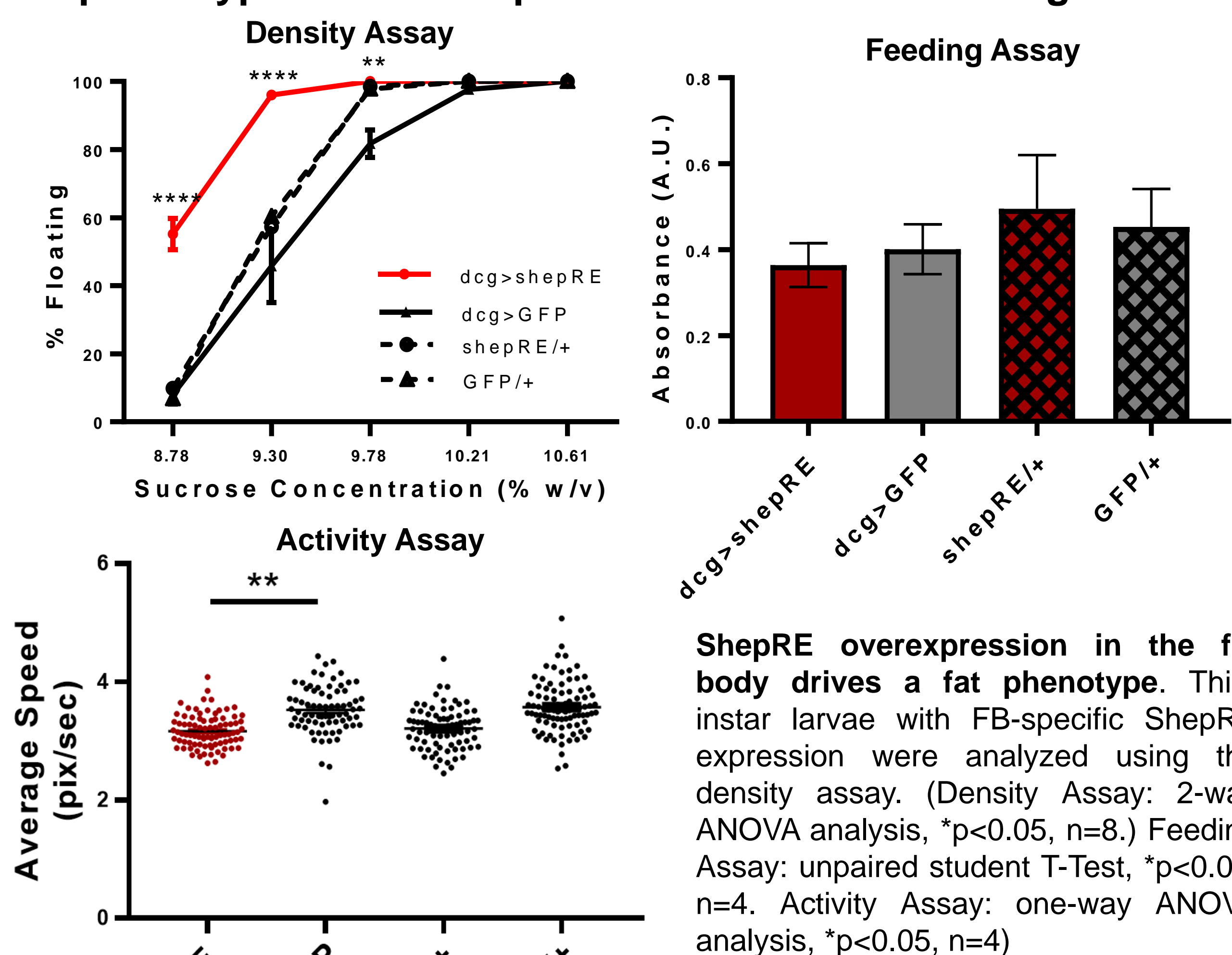
Shep has 8 mRNA and 6 protein isoforms. We hypothesize that fat storage is regulated by one (or more) isoforms. It has two highly conserved RNA Recognition Motifs (RRMs).

Diet modulates FB *shep* isoform expression. *Shep*-PE is the most responsive isoform to diet challenge and expression increases with increased nutrient content.



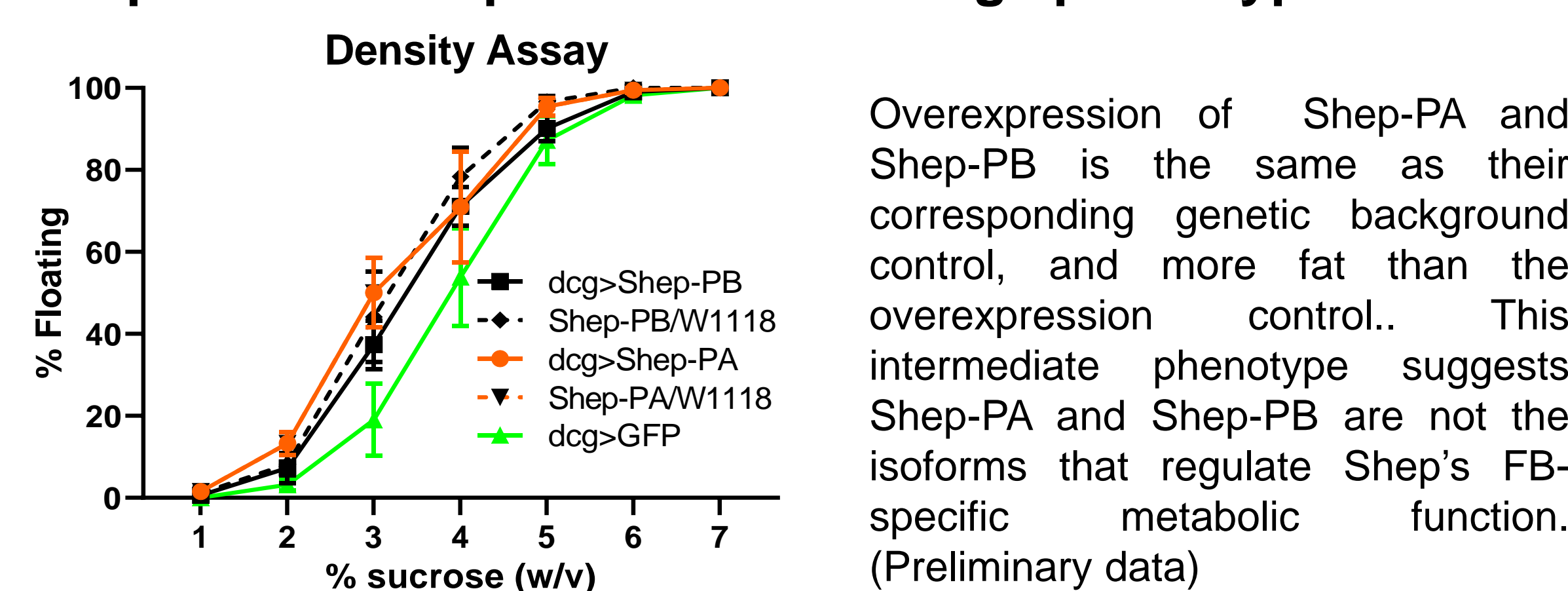
FB *shep* isoform expression changes with diet challenge. RNAseq of dissected fat bodies from w¹¹¹⁸ wandering third-instar larvae reared on Low-Yeast Diet (LYD), Medium-Yeast Diet (MYD), and High-Yeast Diet (HYD). (Two-way ANOVA analysis, n=3, *p<0.05). Western blot of isolated fat bodies from w¹¹¹⁸ wandering third-instar larvae. aShep antibody and size separation identify three broad classes of *Shep* bands: *Shep*-PA/PF, *Shep*-PB, and *Shep*-PE/PH. (Western imaged using Odyssey Li-Cor and analyzed using ImageStudio Lite. 2-Way ANOVA Analysis, n=3).

FB-specific *Shep*RE overexpression induces a fat phenotype that is independent of behavioral changes



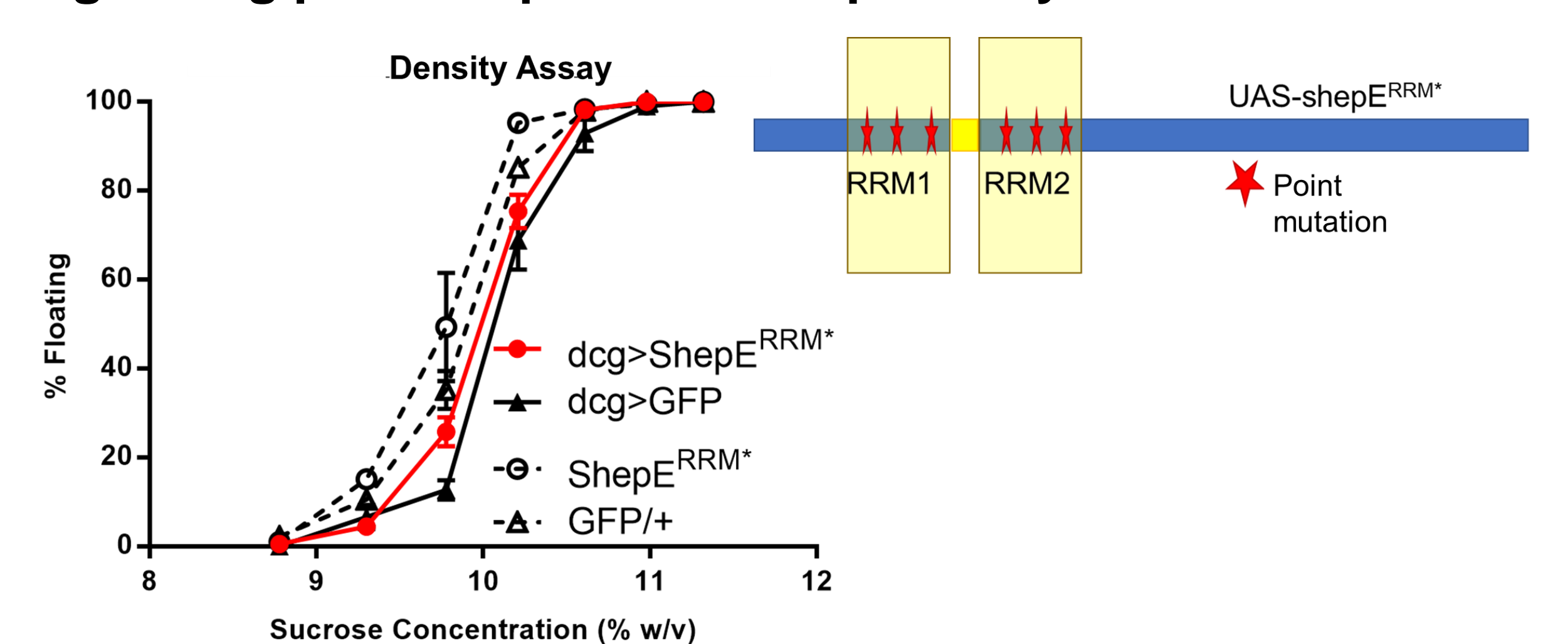
***Shep*RE overexpression in the fat body drives a fat phenotype.** Third instar larvae with FB-specific *Shep*RE expression were analyzed using the density assay. (Density Assay: 2-way ANOVA analysis, *p<0.05, n=8.) Feeding Assay: unpaired student T-Test, *p<0.05, n=4. Activity Assay: one-way ANOVA analysis, *p<0.05, n=4)

Shep-PA and Shep-PB lack fat storage phenotypes

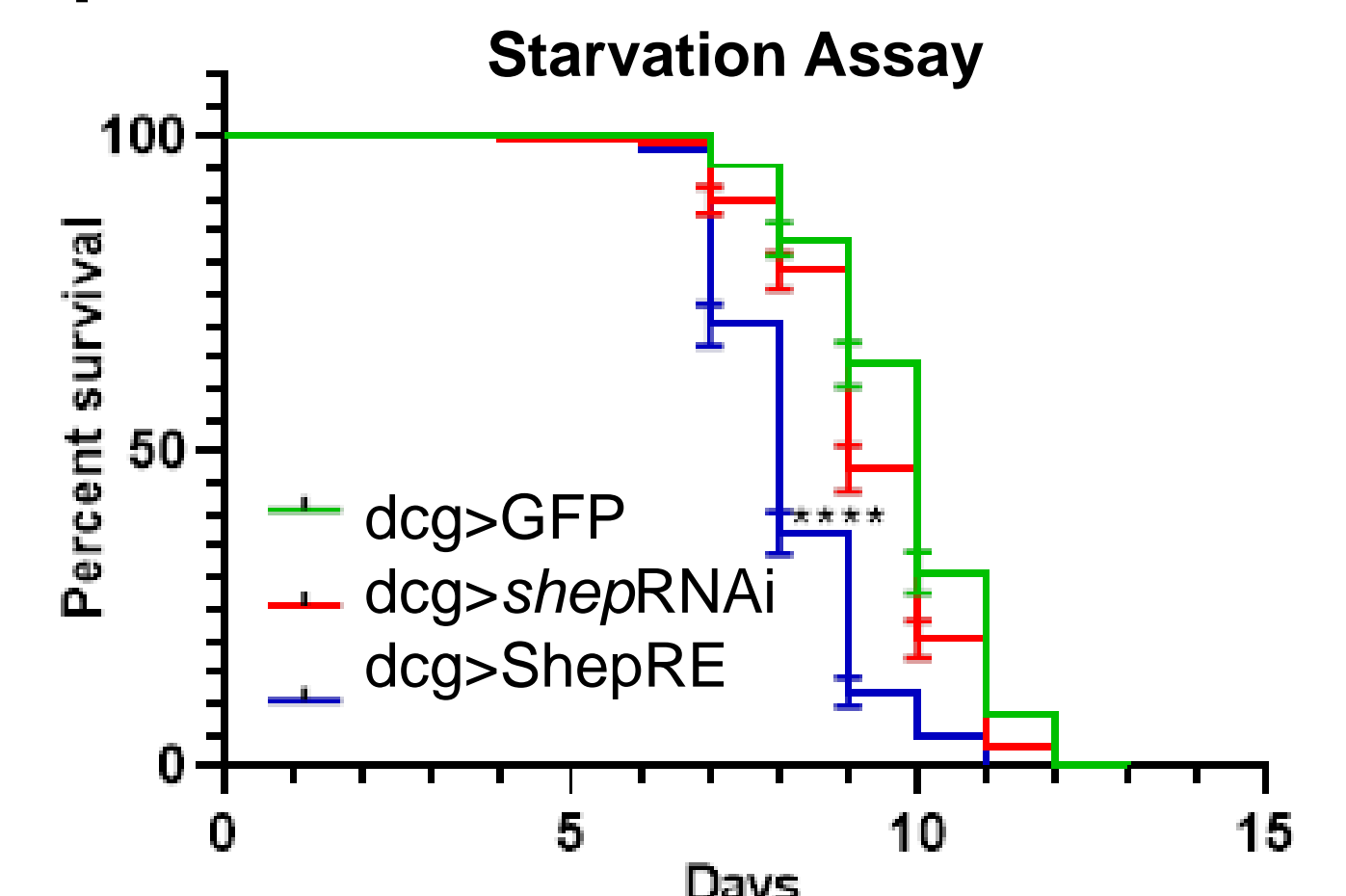


Overexpression of *Shep*-PA and *Shep*-PB is the same as their corresponding genetic background control, and more fat than the overexpression control.. This intermediate phenotype suggests *Shep*-PA and *Shep*-PB are not the isoforms that regulate *Shep*'s FB-specific metabolic function. (Preliminary data)

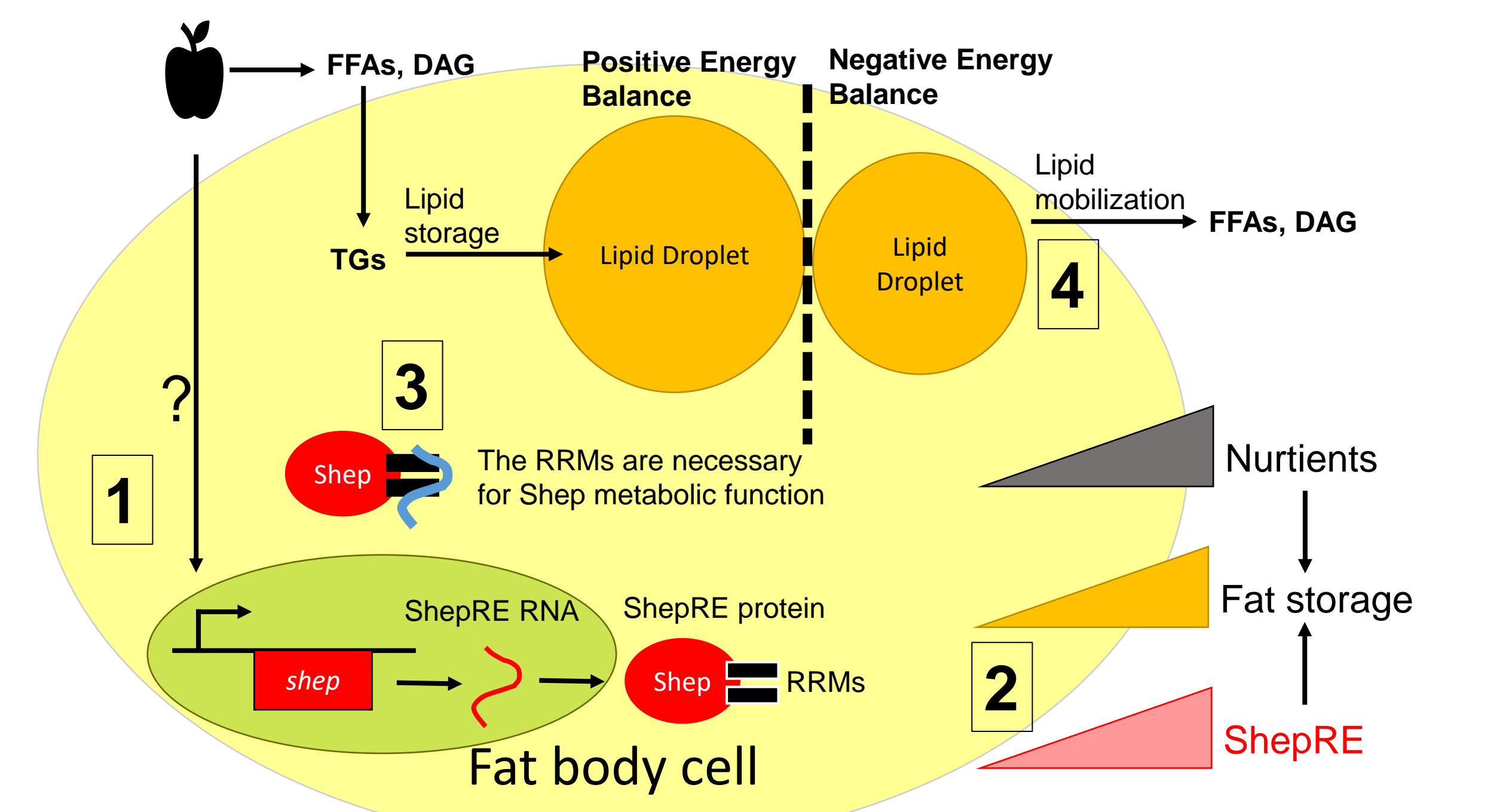
RNA-binding is necessary for *Shep*RE fat storage regulation. We hypothesize that *Shep*RE regulates lipid regulating proteins post-transcriptionally.



ShepRE overexpression makes larvae starvation sensitive



Larvae were kept on a starvation media. *Shep*-RE overexpression dies a mean 20% faster than overexpression control. We hypothesize that this could be due to impaired ability to liberate stored lipids (****p<0.0001, log-rank).



Shep was identified as obesity predisposing gene in a forward genetic screen (Reis, 2010). We showed that FB-specific *shep* depletion caused a lean phenotype. Additionally, *shep* mRNA/protein expression increases as nutrient content of diet increases. The *Shep*RE isoform changes the most upon diet challenge. FB-specific overexpression of *Shep*RE drives a fat phenotype. This phenotype requires the RRM's.

Conclusions on *Shep* function within the fat body:

1. As nutrient content of diet increases, *Shep*RE expression increases
2. *Shep*RE overexpression has a fat phenotype independent of behavior
3. The RRM's are necessary for *Shep*RE fat regulatory function
4. *Shep*RE overexpression makes larvae starvation sensitive, we hypothesize due to impaired ability to liberate stored fat

Acknowledgements and References

We would like to thank Dahong Chen and Elissa Lei (NIDDK, Maryland) for generously sharing their UAS-*Shep* *Drosophila* lines. We would like to thank Dr. Kathleen Beckingham (Rice University) for generously sharing her Rb- α -*Shep* and gp- α -*Shep* antibodies. We would like to acknowledge the University of Colorado Molecular Biology PhD training program, CU-AMC Department of Endocrinology, Diabetes, and Metabolism, and the CU-AMC RNA Bioscience Initiative for their support.

Brooks, A. N., Duff, M. O., May, G., Yang, L., Bolisetty, M., Landolin, J., ... Brenner, S. E. (2015). *Genome Research*, 25(11), 1771–1780. <https://doi.org/10.1101/gr.192518.115>
Chen, D., Gu, T., Pham, T. N., Zachary, M. J., & Hewes, R. S. (2017). <https://doi.org/10.1534/genetics.117.200378>
Matzat, L. H., Dale, R. K., Moshkovich, N., & Lei, E. P. (2012). <https://doi.org/10.1371/journal.pgen.1003069>
Reis, T., Van Gilst, M. R., & Hariharan, I. K. (2010). <https://doi.org/10.1371/journal.pgen.1001206>
Padmanabha, D., & Baker, K. D. (2014). <https://doi.org/10.1016/j.tem.2014.03.011>
Qi, L., et al. Genetic variants at 2q24 are associated with susceptibility to type 2 diabetes. *Hum. Mol. Genet.* **19**, 2706–2715 (2010).
Wu, Y., et al. Down regulation of RNA binding motif, single-stranded interacting protein 3, along with up regulation of nuclear HIF1A correlates with poor prognosis in patients with gastric cancer. *Oncotarget* **8**, 1262–1277 (2017).
Campbell, D. D., et al. Amerind ancestry, socioeconomic status and the genetics of type 2 diabetes in a Colombian population. *PLoS One* **7**, e33570 (2012).