



# Identification of mRNA targets of ETR-1/CELF that regulate Q neuroblast migration in muscle cells using fluorescence-activated cell sorting and RNA-seq



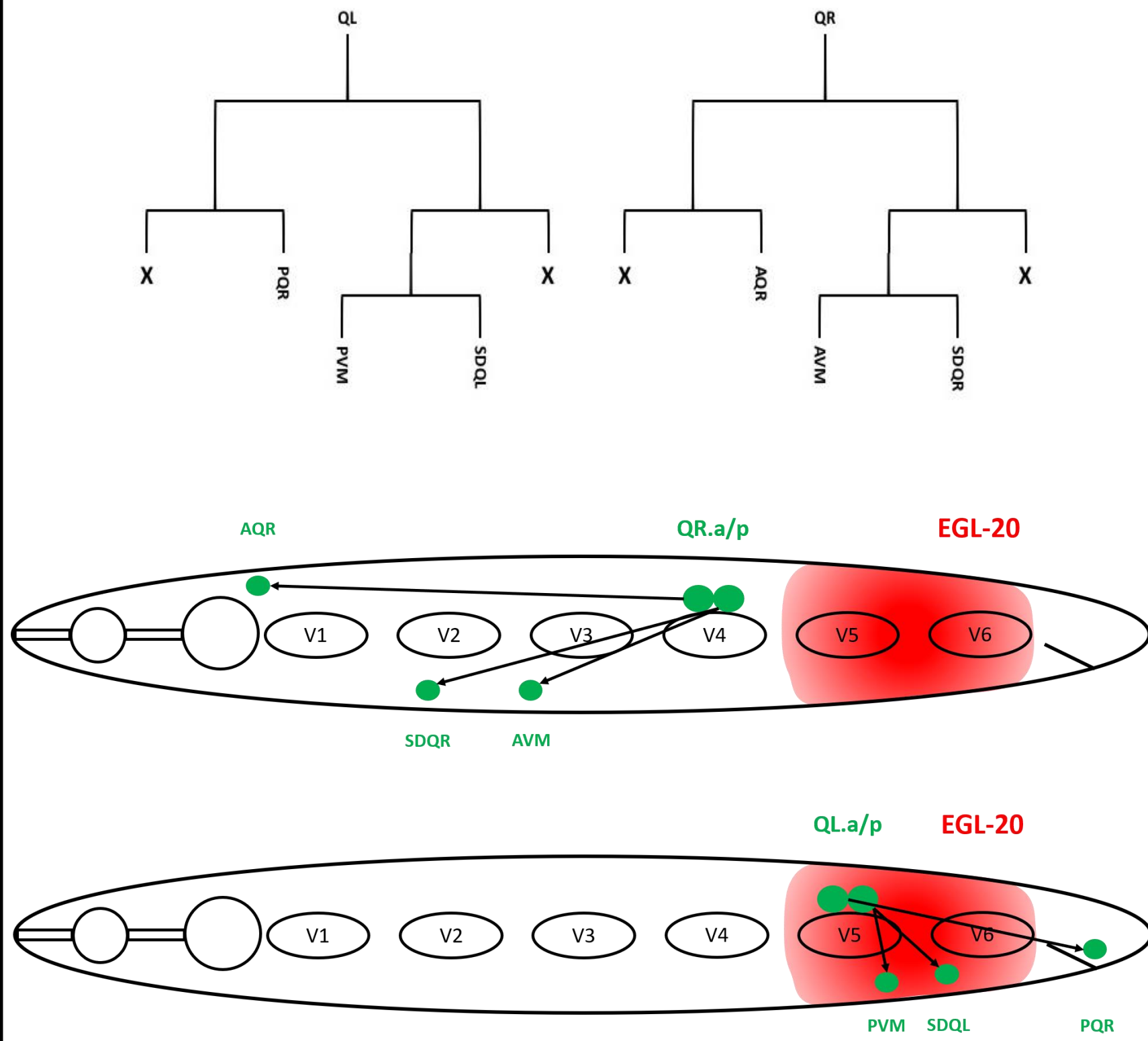
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## Introduction

Migration of neurons is a critical process for the development of the nervous system. *Caenorhabditis elegans* is an excellent model to study cell migration. The Q neuroblasts are bilaterally symmetrical, with QR and its descendants migrating anteriorly, and QL and its descendants on the left migrating posteriorly. These migrations are highly stereotyped. When QL begins its migration towards the posterior it is confronted with a Wnt signal, EGL-20. EGL-20 activates a canonical Wnt pathway, which expresses MAB-5 in QL. MAB-5 is not expressed in QR, which migrates anteriorly. Four other Wnt molecules in *C. elegans* are activated during Q neuroblast migration, CWN-1, CWN-2, LIN-44 and MOM-2. It is clear that Wnt signaling plays a critical role in the migration of the Q neuroblasts during development of the nervous system.

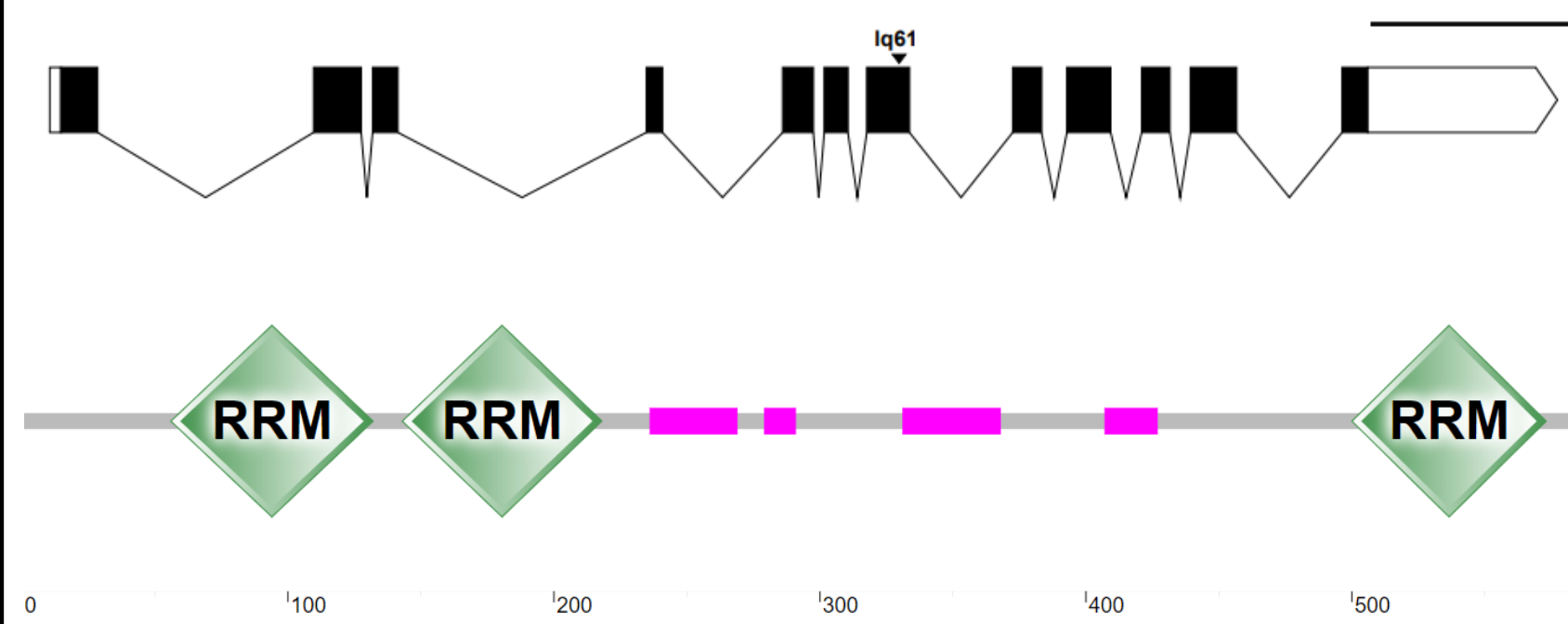
A forward genetic screen identified a mutation in the gene *etr-1*, which caused migration defects in the descendants of QR and QL. *etr-1* is an ELAV-type RNA binding protein, and it is known for its role in the development of muscle. The human homolog for *etr-1*, CUG-binding protein CELF1, has a central role in triplet nucleotide expansion diseases, including myotonic dystrophy type 1. In *C. elegans* mutations in *etr-1* usually result in severe muscle disorganization, paralysis at the two-fold stage, and lethality. We have isolated a viable mutation from our screen which results in a premature stop codon in an alternatively spliced exon. We have also generated another allele with CRISPR mediated knockdown, with a frameshift mutation in the same alternatively spliced exon.

## Q Neuroblasts are Bilaterally Symmetrical

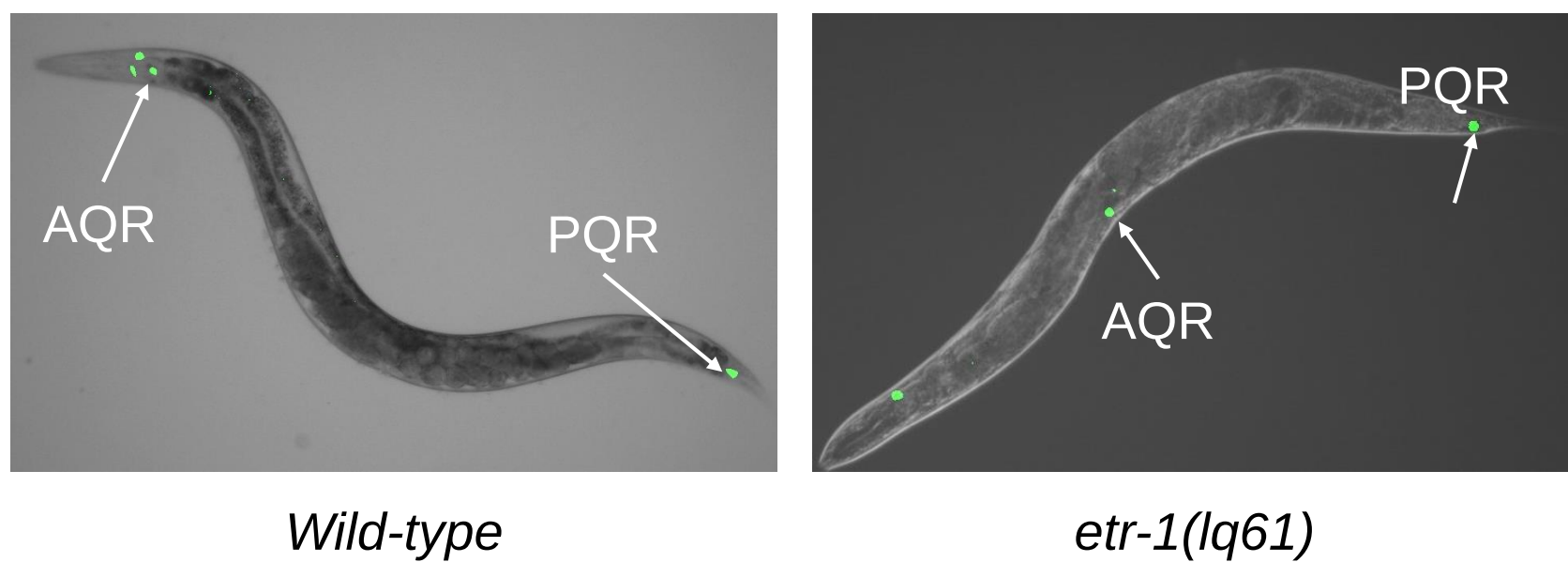


## *etr-1* encodes an RNA-binding protein

- lq61* corresponds to a premature stop codon in an alternatively spliced exon
- Total of 84 different isoforms identified, *lq61* affects only ten. Genomic sequence is depicted below, showing all exons
- Protein is characterized by three RNA-recognition motifs



## *etr-1* causes AQR/PQR defects

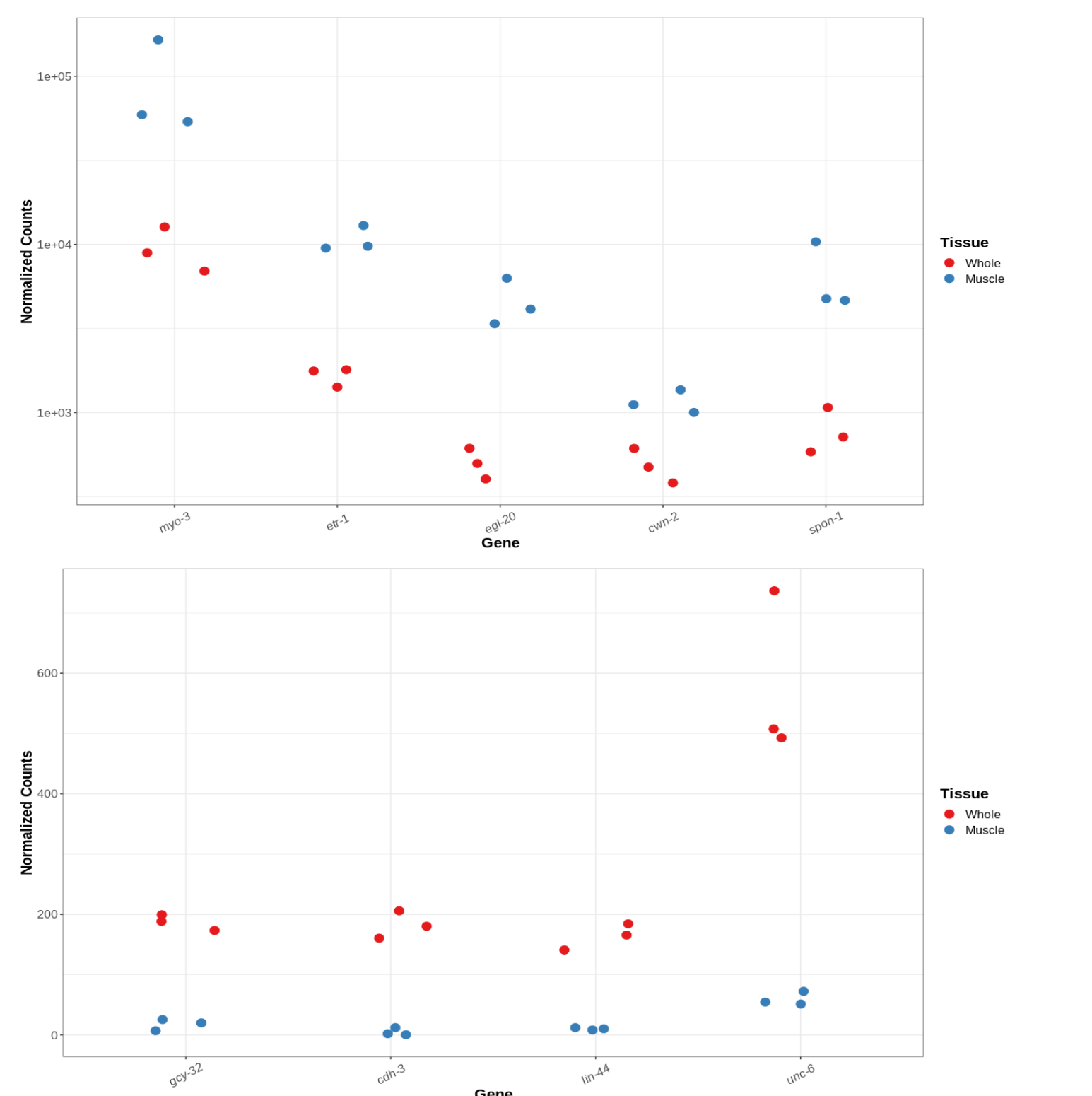


| Genotype                          | AQR |    |   |   |   |     | PQR |   |   |    |     |     | p      |
|-----------------------------------|-----|----|---|---|---|-----|-----|---|---|----|-----|-----|--------|
|                                   | 1   | 2  | 3 | 4 | 5 | n   | 1   | 2 | 3 | 4  | 5   | n   |        |
| Wild-type                         | 100 | 0  | 0 | 0 | 0 | 100 | 0   | 0 | 0 | 0  | 100 | 100 |        |
| <i>etr-1(lq61)</i>                | 85  | 12 | 2 | 0 | 1 | 100 | 3   | 5 | 4 | 0  | 88  | 100 | <0.001 |
| <i>etr-1(lq133)</i>               | 75  | 19 | 3 | 3 | 0 | 100 | 1   | 1 | 5 | 10 | 82  | 100 |        |
| <i>etr-1(lq61);lqEx[etr-1(+)]</i> | 97  | 2  | 0 | 0 | 1 | 100 | 0   | 0 | 0 | 0  | 100 | 100 | <0.01  |

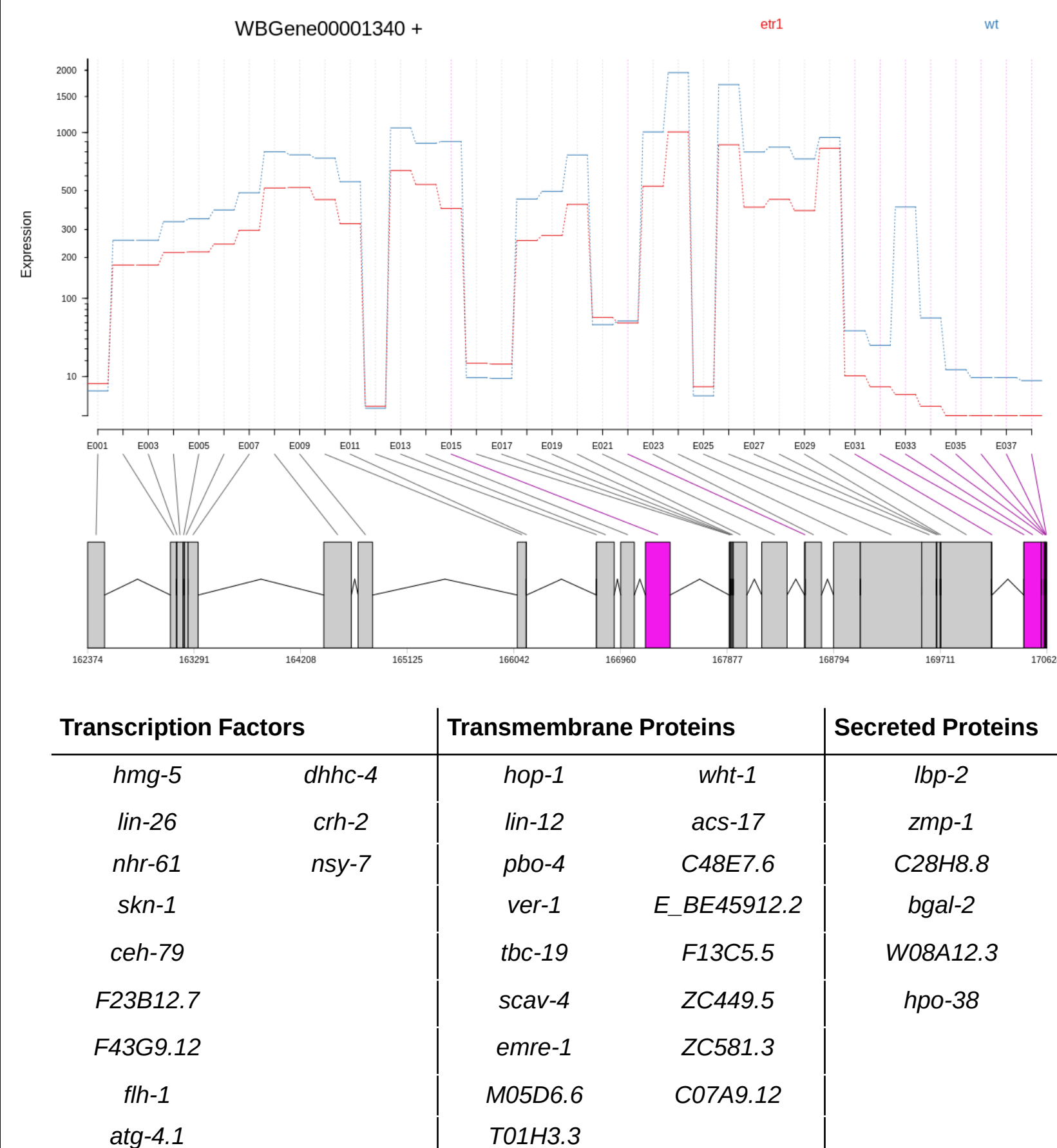
## *etr-1* acts non-autonomously in muscle cells

| Genotype                      | AQR |    |   |   |   |        | PQR |   |   |    |    |        | p |
|-------------------------------|-----|----|---|---|---|--------|-----|---|---|----|----|--------|---|
|                               | 1   | 2  | 3 | 4 | 5 | p      | 1   | 2 | 3 | 4  | 5  | p      |   |
| <i>Pegl-17::etr-1(+)</i>      |     |    |   |   |   |        |     |   |   |    |    |        |   |
| <i>etr-1(lq61)</i>            | 85  | 12 | 2 | 0 | 1 |        | 3   | 5 | 4 | 0  | 88 |        |   |
| <i>etr-1(lq61) lqEx912</i>    | 85  | 11 | 0 | 2 | 1 |        | 0   | 0 | 0 | 9  | 91 |        |   |
| <i>etr-1(lq61) no lqEx912</i> | 84  | 14 | 0 | 0 | 2 | 1      | 0   | 0 | 0 | 10 | 90 | 1      |   |
| <i>etr-1(lq61) lqEx913</i>    | 88  | 7  | 1 | 2 | 2 |        | 0   | 0 | 0 | 11 | 89 |        |   |
| <i>etr-1(lq61) no lqEx913</i> | 85  | 10 | 1 | 1 | 3 | 1      | 0   | 0 | 0 | 8  | 92 | 1      |   |
| <i>etr-1(lq61) lqEx914</i>    | 87  | 10 | 2 | 1 | 0 |        | 1   | 0 | 5 | 4  | 90 |        |   |
| <i>etr-1(lq61) no lqEx914</i> | 83  | 13 | 2 | 2 | 0 | 0.5530 | 1   | 1 | 3 | 6  | 89 | 1      |   |
| <i>Pmyo-3::etr-1(+)</i>       |     |    |   |   |   |        |     |   |   |    |    |        |   |
| <i>etr-1(lq61)</i>            | 85  | 12 | 2 | 0 | 1 |        | 3   | 5 | 4 | 0  | 88 |        |   |
| <i>etr-1(lq61) lqEx944</i>    | 100 | 0  | 0 | 0 | 0 | <0.001 | 2   | 0 | 0 | 0  | 98 |        |   |
| <i>etr-1(lq61) no lqEx944</i> | 70  | 22 | 5 | 1 | 2 |        | 2   | 1 | 3 | 7  | 87 | 0.0055 |   |
| <i>etr-1(lq61) lqEx945</i>    | 86  | 14 | 0 | 0 | 0 | 0.0509 | 0   | 0 | 0 | 2  | 98 |        |   |
| <i>etr-1(lq61) no lqEx945</i> | 74  | 24 | 1 | 1 | 0 |        | 4   | 3 | 0 | 5  | 88 | 0.0101 |   |
| <i>etr-1(lq61) lqEx946</i>    | 94  | 6  | 0 | 0 | 0 | 0.0092 | 0   | 0 | 0 | 1  | 99 |        |   |
| <i>etr-1(lq61) no lqEx946</i> | 81  | 14 | 2 | 1 | 2 |        | 0   | 0 | 0 | 1  | 99 | 1.000  |   |
| <i>etr-1(lq61) lqEx947</i>    | 97  | 3  | 0 | 0 | 0 | <0.001 | 1   | 0 | 0 | 0  | 99 |        |   |
| <i>etr-1(lq61) no lqEx947</i> | 79  | 14 | 6 | 0 | 1 |        | 1   | 1 | 0 | 4  | 94 | 0.0649 |   |

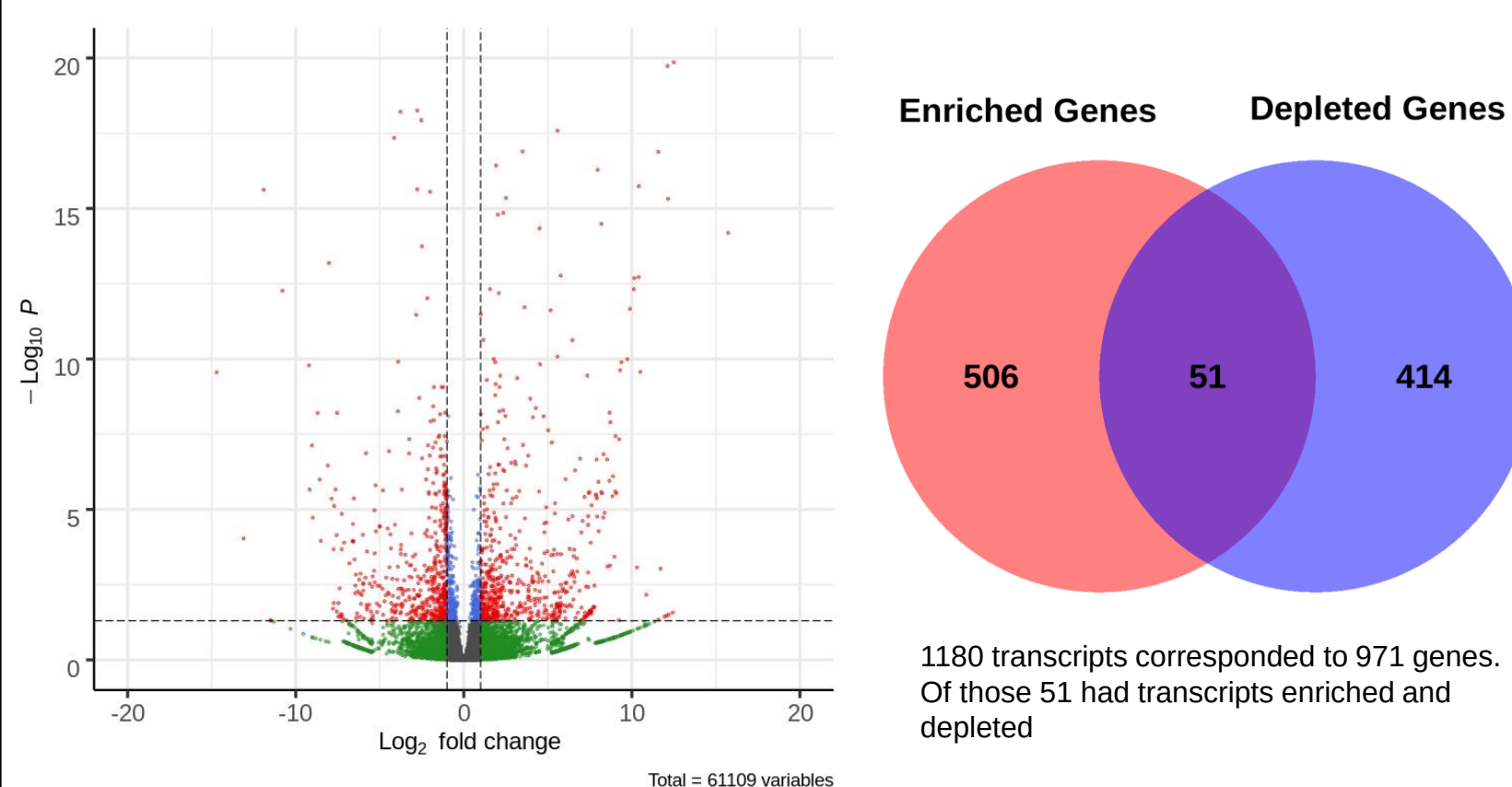
## Normalized counts of genes to show proper enrichment and depletion in wild-type muscles



## 244 genes had differential exon usage when comparing wild-type muscle to *etr-1* muscle



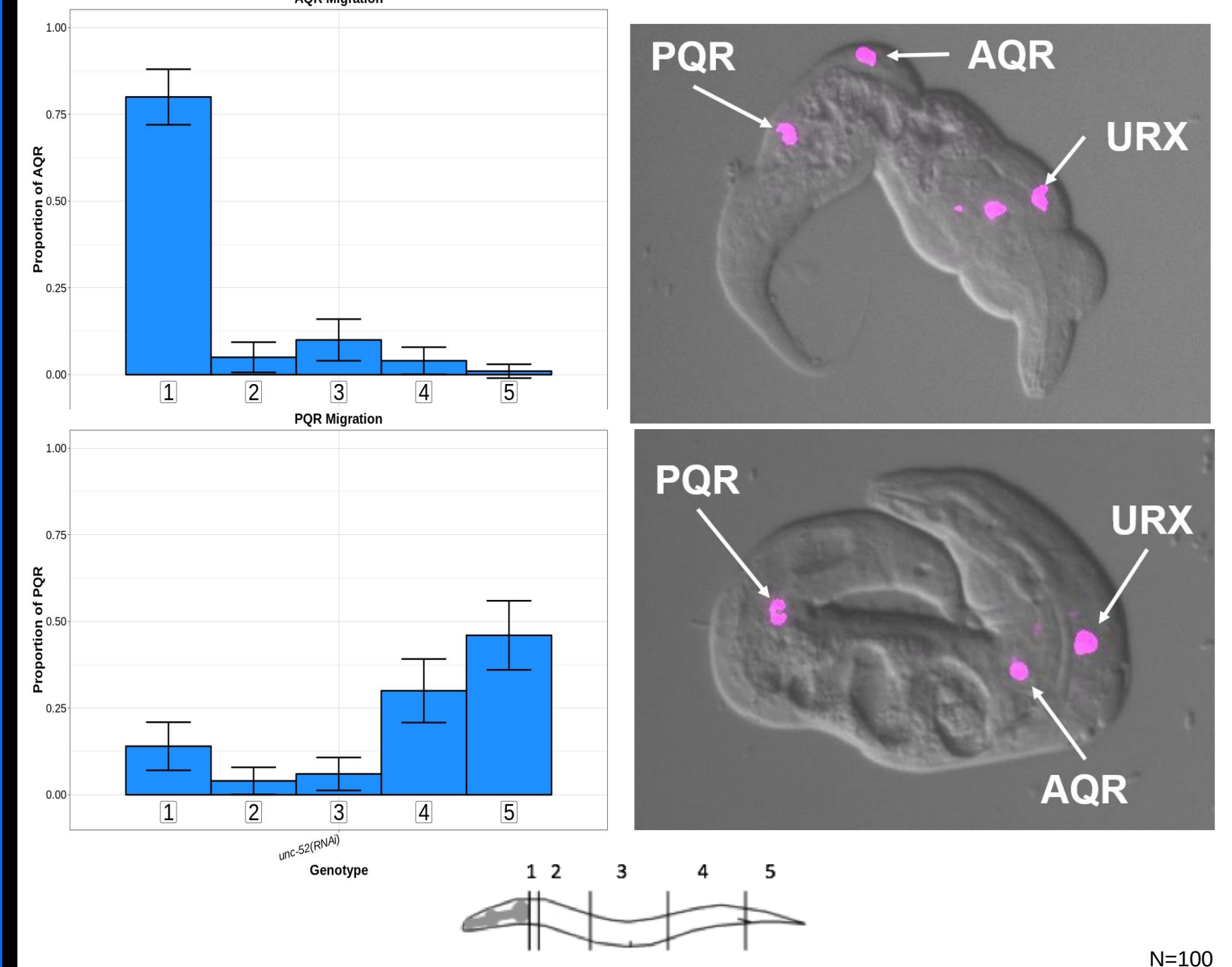
## 1180 transcripts were differentially expressed when comparing wild-type to *etr-1* muscles



## 102 genes were identified in both transcript and exon usage analyses

|               |               |                   |                 |                |
|---------------|---------------|-------------------|-----------------|----------------|
| <i>atn-1</i>  | <i>mrp-1</i>  | <i>unc-73</i>     | <i>cpna-2</i>   | <i>ZK370.4</i> |
| <i>avr-14</i> | <i>nhr-61</i> | <i>unc-87</i>     | <i>whi-1</i>    | <i>nsy-7</i>   |
| <i>ccr-4</i>  | <i>ntl-2</i>  | <i>unc-89</i>     | <i>fln-2</i>    |                |
| <i>ced-1</i>  | <i>num-1</i>  | <i>unc-96</i>     | <i>crh-2</i>    |                |
| <i>ctn-1</i>  | <i>ketn-1</i> | <i>ver-1</i>      | <i>C27H5.2</i>  |                |
| <i>deb-1</i>  | <i>ret-1</i>  | <i>zmp-1</i>      | <i>pxl-1</i>    |                |
| <i>dur-1</i>  | <i>rme-1</i>  | <i>alk-1</i>      | <i>tag-250</i>  |                |
| <i>alp-1</i>  | <i>rpl-19</i> | <i>C06B3.6</i>    | <i>acs-17</i>   |                |
| <i>egl-10</i> | <i>sli-1</i>  | <i>ceh-79</i>     | <i>fhod-1</i>   |                |
| <i>etr-1</i>  | <i>slo-1</i>  | <i>tbc-19</i>     | <i>C56G2.9</i>  |                |
| <i>gcy-31</i> | <i>srp-7</i>  | <i>pde-3</i>      | <i>F13C5.5</i>  |                |
| <i>gld-2</i>  | <i>lim-9</i>  | <i>shw-1</i>      | <i>F20A1.6</i>  |                |
| <i>gon-1</i>  | <i>ttn-1</i>  | <i>F23B12.7</i>   | <i>dip-2</i>    |                |
| <i>hsp-4</i>  | <i>dgk-4</i>  | <i>pole-1</i>     | <i>F44E7.4</i>  |                |
| <i>kin-1</i>  | <i>cpna-1</i> | <i>mlcd-1</i>     | <i>F46F11.1</i> |                |
| <i>lea-1</i>  | <i>tns-1</i>  | <i>F40F8.5</i>    | <i>F48E3.8</i>  |                |
| <i>lec-2</i>  | <i>tax-6</i>  | <i>F43G9.12</i>   | <i>K09F6.10</i> |                |
| <i>lec-3</i>  | <i>tmd-2</i>  | <i>F49E2.5</i>    | <i>trx-3</i>    |                |
| <i>lev-11</i> | <i>tnt-3</i>  | <i>yif-1</i>      | <i>M01H9.3</i>  |                |
| <i>lin-45</i> | <i>tsp-17</i> | <i>magi-1</i>     | <i>R11G1.6</i>  |                |
| <i>lip-1</i>  | <i>unc-32</i> | <i>nep-21</i>     | <i>T23E7.2</i>  |                |
| <i>lsm-5</i>  | <i>unc-36</i> | <i>T28F4.5</i>    | <i>zig-11</i>   |                |
| <i>mai-1</i>  | <i>unc-52</i> | <i>grdn-1</i>     | <i>fln-1</i>    |                |
| <i>mel-11</i> | <i>unc-64</i> | <i>Y105E8A.25</i> | <i>him-19</i>   |                |
| <i>mlp-1</i>  | <i>unc-68</i> | <i>nit-1</i>      | <i>ZC449.5</i>  |                |

## *unc-52(RNAi)* results in severe AQR and PQR migration defects



## Most AQR and PQR migration defects are minor

| Genotype             | AQR |    |   |   |   | PQR |   |   |   |     | n   |
|----------------------|-----|----|---|---|---|-----|---|---|---|-----|-----|
|                      | 1   | 2  | 3 | 4 | 5 | 1   | 2 | 3 | 4 | 5   |     |
| <i>mel-11(RNAi)</i>  | 94  | 6  | 0 | 0 | 0 | 0   | 0 | 0 | 0 | 100 | 100 |
| <i>unc-89(RNAi)</i>  | 89  | 11 | 0 | 0 | 0 | 0   | 0 | 0 | 0 | 100 | 100 |
| <i>ced-1(RNAi)</i>   | 92  | 8  | 0 | 0 | 0 | 0   | 0 | 0 | 1 | 99  | 100 |
| <i>atn-1(RNAi)</i>   | 92  | 8  | 0 | 0 | 0 | 0   | 0 | 0 | 0 | 100 | 100 |
| <i>unc-32(RNAi)</i>  | 98  | 2  | 0 | 0 | 0 | 0   | 0 | 0 | 0 | 100 | 100 |
| <i>lea-1(RNAi)</i>   | 89  | 11 | 0 | 0 | 0 | 0   | 0 | 0 | 0 | 100 | 100 |
| <i>unc-87(RNAi)</i>  | 96  | 3  | 1 | 0 | 0 | 0   | 0 | 0 | 0 | 100 | 100 |
| <i>tnt-3(RNAi)</i>   | 95  | 5  | 0 | 0 | 0 | 0   | 0 | 0 | 0 | 100 | 100 |
| <i>hmg-5(RNAi)</i>   | 80  | 20 | 0 | 0 | 0 | 0   | 0 | 0 | 0 | 100 | 100 |
| <i>ver-1(RNAi)</i>   | 78  | 22 | 0 | 0 | 0 | 0   | 0 | 0 | 0 | 100 | 100 |
| <i>lin-45(RNAi)</i>  | 85  | 15 | 0 | 0 | 0 | 0   | 0 | 0 | 0 | 100 | 100 |
| <i>ceh-79(RNAi)</i>  | 92  | 8  | 0 | 0 | 0 | 0   | 0 | 0 | 0 | 100 | 100 |
| <i>mig-22(RNAi)</i>  | 88  | 12 | 0 | 0 | 0 | 0   | 0 | 0 | 3 | 97  | 100 |
| <i>sop-2(RNAi)</i>   | 87  | 12 | 1 | 0 | 0 | 0   | 0 | 0 | 1 | 99  | 100 |
| <i>unc-112(RNAi)</i> | 98  | 2  | 0 | 0 | 0 | 0   | 0 | 0 | 1 | 99  | 100 |
| <i>unc-71(RNAi)</i>  | 91  | 4  | 0 | 0 | 5 | 5   | 0 | 0 | 0 | 95  | 100 |
| <i>noca-1(RNAi)</i>  | 87  | 12 | 1 | 0 | 0 | 0   | 0 | 0 | 0 | 100 | 100 |
| <i>ani-1(RNAi)</i>   | 89  | 11 | 0 | 0 | 0 | 0   | 0 | 0 | 2 | 98  | 100 |

## Conclusions and future directions

- etr-1* causes defects in the migration of the Q neuroblast descendants
- etr-1* acts in the muscle cells
  - Pmyo-3::etr-1* rescues defects
- We have identified 244 potential genes that are alternatively spliced by ETR-1
- We have identified 1180 transcripts that are potential targets of ETR-1
- RNAi knockdown of *unc-52* displays severe AQR and PQR migration defects
- RNAi knockdown of most genes identified resulted in minor AQR and PQR defects

## Acknowledgements

### Lundquist lab members

Dr. Erik Lundquist  
Eric Struckhoff  
Dr. Vitoria Paolillo  
Snehal Mahadik  
Kelsey Ferguson  
Aubrie Stricker  
Sierra Mortimer  
Angela Lang

### Dr. Brian Ackley and lab

Funding:  
R21 NS100483  
P20 GM103638

