



Deciphering Roles of Bloom Syndrome Helicase (Blm) in Genome Stability

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BACKGROUND AND INTRODUCTION

(1) Mitotic Crossovers (mitCOs) resulting from Homologous Recombination (HR) repair of DNA are highly detrimental to cells, leading to potential loss of heterozygosity (LOH), genome instability, and cancer^{1,2}.

(2) Bloom Syndrome Helicase (*Drosophila*: *Blm*; Human: *BLM*) has been shown to cause **increased** spontaneous mitotic crossover rates when absent in the *Drosophila* male germline, mammalian cells, and Human Bloom Syndrome cells³⁻⁷.

(3) Accurate interpretation of mitCO mechanisms in multicellular organisms was previously difficult due to obfuscation of heteroduplex DNA tracts via canonical and short-patch mismatch repair (cMMR and spMMR). Complete MMR knockout (KO) has **only** been achieved in *Drosophila*, leading to a new model for meiotic crossovers⁸.

(4) Gain in copy number of BLM has been observed in several cancers, suggesting potential alternative roles for Blm in these cancers.

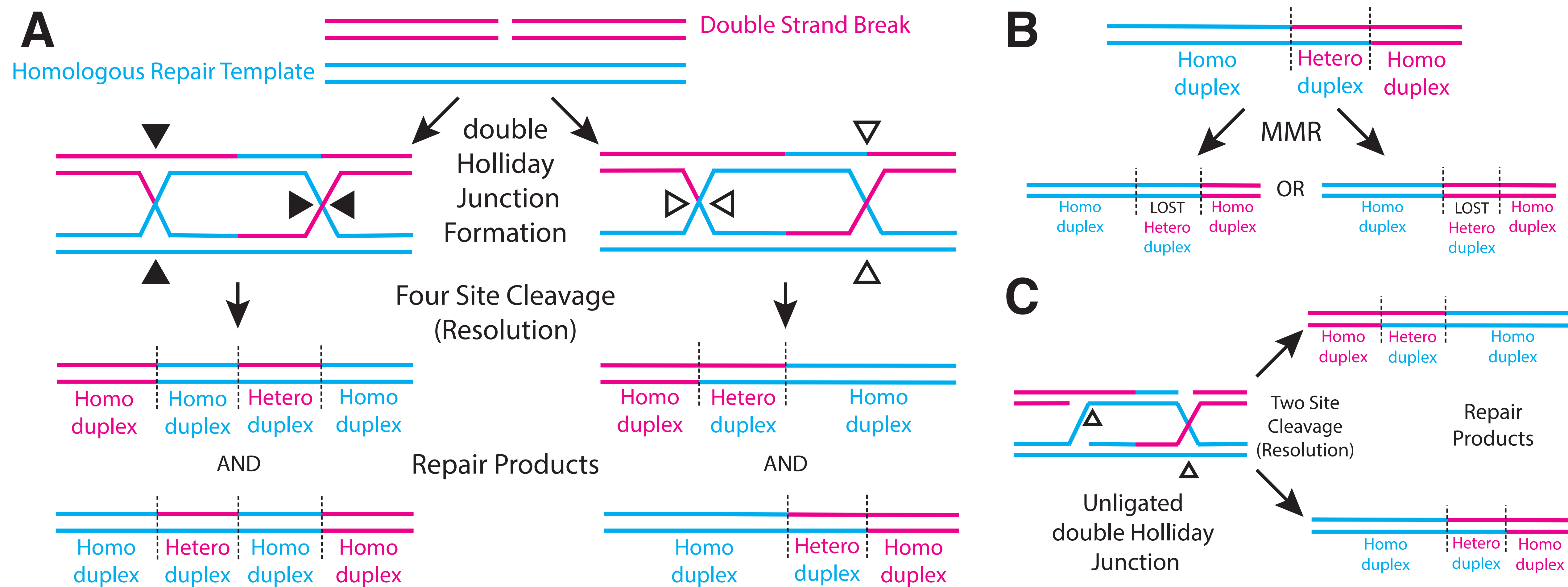


Figure 1. (A) Double Strand Break (DSB) repair mediated by Homologous Recombination (HR) repair. Double Holliday Junction (dHJ) intermediates are one model for how HR repairs DSBs. Differing dHJ cleavages at four sites (filled vs. open arrowheads) result in distinct hetero- and homoduplex DNA arrangements in repair products. Some products have four distinct DNA duplexes surrounding the repaired DSB (left), while others have three (right). These arrangements help determine where a DSB originated and how it was repaired. **(B)** Mismatch Repair (MMR) converts informative heteroduplex DNA to homoduplex DNA, making finding where a DSB originated and how it was repaired difficult to interpret depending on how the conversion occurs. This is done via canonical and short-patch (cMMR and spMMR) pathways. **(C)** A meiotic crossover (CO) model suggests use of an unligated dHJ and two site cleavage (resolution) rather than four site cleavage based on KO of MMR and the meiotic CO products observed⁸. Such a model could be applicable to mitotic COs (mitCOs).

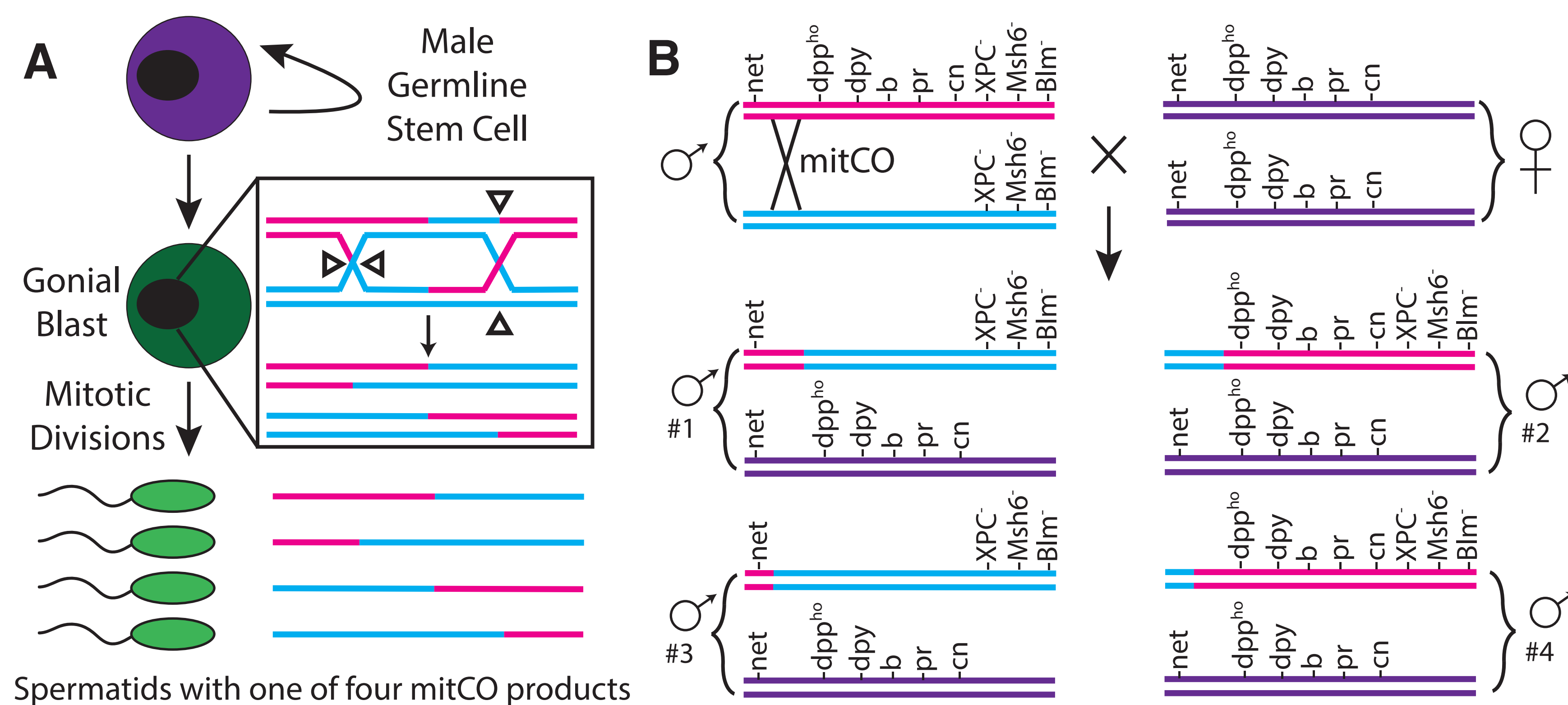


Figure 2. (A) Depiction of *Drosophila* male germline divisions. A mitCO during the Gonialblast division (callout box) results in spermatids with one of four products. **(B)** By mating males heterozygous for net-cn and homozygous for spMMR and cMMR (XPC and Msh6, respectively) knockout (KO) and Blm KO to net-cn females, mitCOs (e.g. between net and dpp^{ho}) are recovered by screening male progeny for reciprocal product arrangements indicated by net, dpp^{ho}, dpy, b, pr, and cn recessive markers (e.g. #1-4). Precise mitCO sites are then mapped via Illumina or Oxford Nanopore whole genome sequencing of one male, followed by Sanger sequencing to map remaining reciprocal products.

HYPOTHESES

(1) Resolution of double Holliday Junctions (dHJ) will prevail as the mechanism for Blm KO spontaneous mitCOs.

(2) Alternative and/or secondary mechanisms may apply to mitCOs, such as one described for meiosis⁸.

(3) Blm overexpression will not cause overgrowth or tumor-like phenotypes in normal epithelia.

(4) Blm gain of copy number in certain cancers will promote growth through alleviation of replication fork damage, and prevent complete collapse of genome stability through HR.

PRELIMINARY DATA AND RESULTS

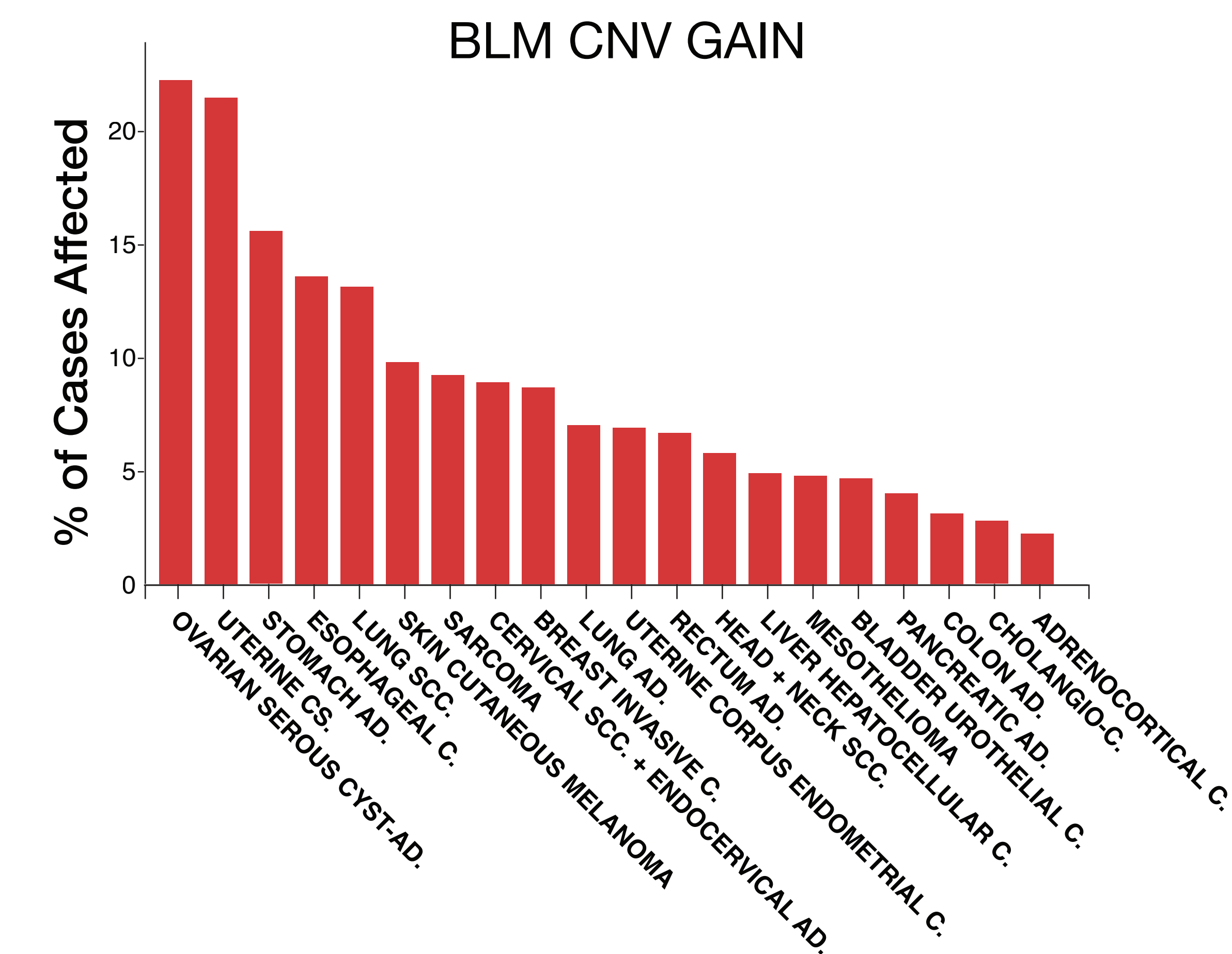


Figure 3. Percentages of Various Cancers Affected by BLM Copy Number Variation (CNV) Gain. While BLM has been previously implicated in both Bloom Syndrome and other cancers by a loss of function, recent data from The Cancer Genome Atlas (TCGA) instead show many cancers gain in BLM copy number, suggesting a potential alternative role for BLM in these cancers.

CONCLUSIONS

(1) Blm overexpression in *Drosophila* imaginal eye discs reduces the amount of DSBs, indicating that copy number gain (and presumably overexpressed) *BLM* could reduce the number of DSBs in cancer to promote basal genomic stability and persistence.

CITATIONS AND ACKNOWLEDGEMENTS

- Chatterjee, N. & Walker, G. C. Mechanisms of DNA damage, repair, and mutagenesis. *Environ Mol Mutagen* 58, 235-263, doi:10.1002/em.22087 (2017).
- Mohaghegh, P. & Hickson, I. D. DNA helicase deficiencies associated with cancer predisposition and premature ageing disorders. *Hum Mol Genet* 10, 741-746, doi:10.1093/hmg/10.7.741 (2001).
- Chaganti, R. S., Schonberg, S. & German, J. A manyfold increase in sister chromatid exchanges in Bloom's syndrome lymphocytes. *Proc Natl Acad Sci U S A* 71, 4508-4512, doi:10.1073/pnas.71.11.4508 (1974).
- German, J. Cytological Evidence for Crossing-over in Vitro in Human Lymphoid Cells. *Science* 144, 298-301, doi:10.1126/science.144.3616.298 (1964).
- LaRocque, J. R. et al. Interhomolog recombination and loss of heterozygosity in wild-type and Bloom syndrome helicase (BLM)-deficient mammalian cells. *Proc Natl Acad Sci U S A* 108, 11971-11976, doi:10.1073/pnas.1104421108 (2011).
- Schroeder, T. M. Sister chromatid exchanges and chromatid interchanges in bloom's syndrome. *Humangenetik* 30, 317-323, doi:10.1007/bf00275144 (1975).
- Suzuki, T., Yasui, M. & Honma, M. Mutator Phenotype and DNA Double-Strand Break Repair in BLM Helicase-Deficient Human Cells. *Mol Cell Biol* 36, 2877-2889, doi:10.1128/MCB.00443-16 (2016).
- Crown, K. N., McMahan, S. & Sekelsky, J. Eliminating both canonical and short-patch mismatch repair in *Drosophila melanogaster* suggests a new meiotic recombination model. *PLoS Genet* 10, e1004583, doi:10.1371/journal.pgen.1004583 (2014)

This work was supported by National Institutes of Health (NIGMS and NCI) grants R35GM118127 and T32CA009156 (Integrated Training in Cancer Model Systems)

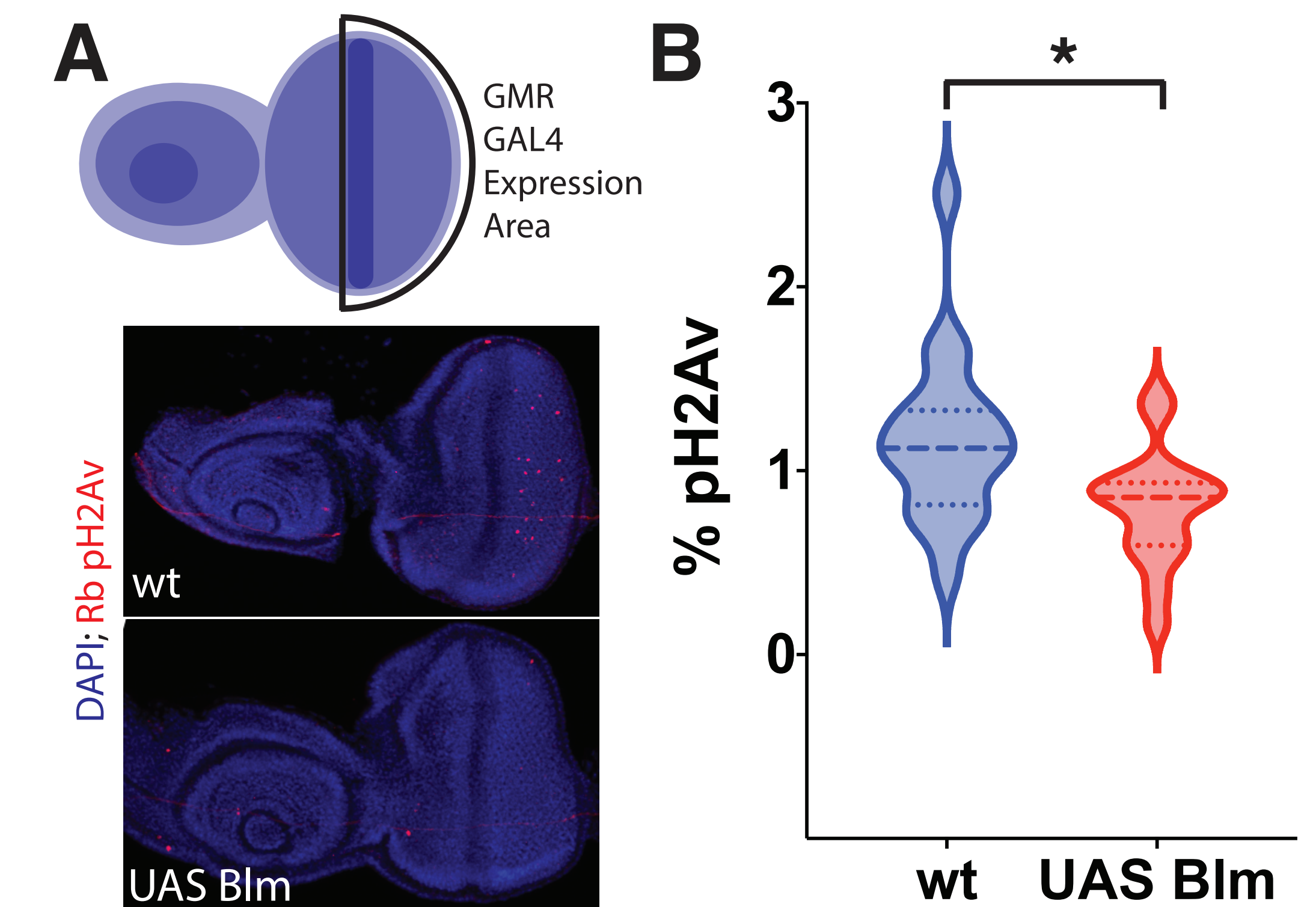


Figure 4. (A) Schematic of GMR GAL4 driver expression area and representative images of wt and UAS Blm (Blm overexpression) *Drosophila* imaginal eye disc epithelia. GMR drives expression to the right of the second mitotic wave cells (dark band) in the larger eye disc, indicated by the black semi-oval outline. Representative images of wt and UAS Blm discs stained with DAPI (blue) and with the DSB marker rabbit (Rb) α -pH2Av (*Drosophila* H2A.X; red). **(B)** Violin plot of percent pH2Av signal in GMR GAL4 expression area shows UAS Blm decreases DSBs compared to wt (*p < 0.05, unpaired t-test). Dashed line = median; dotted line = 1st and 3rd quartiles.