

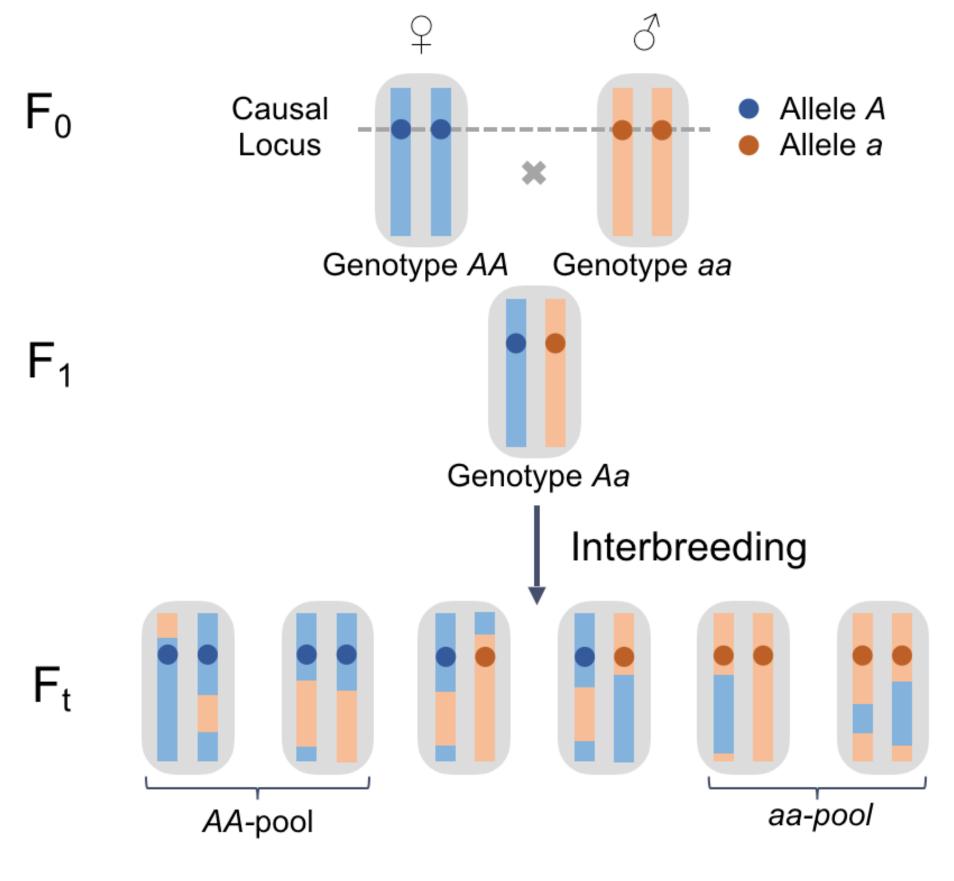
Predicting the Genomic Resolution of Bulk Segregant Analysis

Abstract

Ancestry breakpoint: the point where ancestry changes along a chromosome, e.g. the Bulk segregant analysis (BSA) is a genetic mapping technique for ancestry from the AA strain in blue switches to the aa strain in orange, or vice versa. identifying the loci that underlie phenotypic variation. The basic principle of this method is to select two pools of individuals from **D**: the distance to the closest ancestry breakpoint downstream of the QTL, observed the opposing tails of the phenotypic distribution for the trait of among all chromosomes in the sample. interest. These pools are then each sequenced and scanned for ▶●---●●---●●---●●---● alleles that show characteristically diverged frequencies between the pools, indicating that they could be responsible for the observed trait differences. BSA has already been successfully The distance **D** to the closest ancestry breakpoint located downstream of the QTL in a applied for the mapping of quantitative trait loci (QTLs) in organisms ranging from yeast to crops. However, these studies sample of 2s gametes from the F_1 (representing a sample of s diploid individuals from the F₂) will then be an exponential random variable with cumulative density function: have typically suffered from rather low genomic resolution, and we still lack a detailed understanding of how this resolution is $P(D \le d) = 1 - e^{-2rsd}$, with $E[D] = \frac{1}{2rs}$. [1] affected by experimental parameters. Here, we use coalescence theory to derive analytical results for the expected mapping resolution of BSA. We first show that in an idealized population without genetic drift the expected mapping resolution is inversely proportional to the recombination rate, the number of generations of interbreeding, and the number of genomes sampled, as intuitively expected. In a finite population, coalescence events in the genealogy of the sample reduce the number of potentially informative recombination events during interbreeding, and thus the achievable mapping resolution. This is incorporated in our theory by introducing an effective population size parameter, specified by the pairwise coalescence rate in the interbreeding population. We show that the mapping resolution predicted by our theory is in excellent accordance with numerical simulations. Our framework can enable researchers to assess the expected power of a given BSA experiment, and to test how experimental setup could be tuned to optimize mapping resolution.

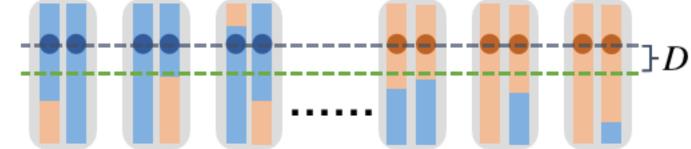
Bulk Segregant Analysis (BSA)

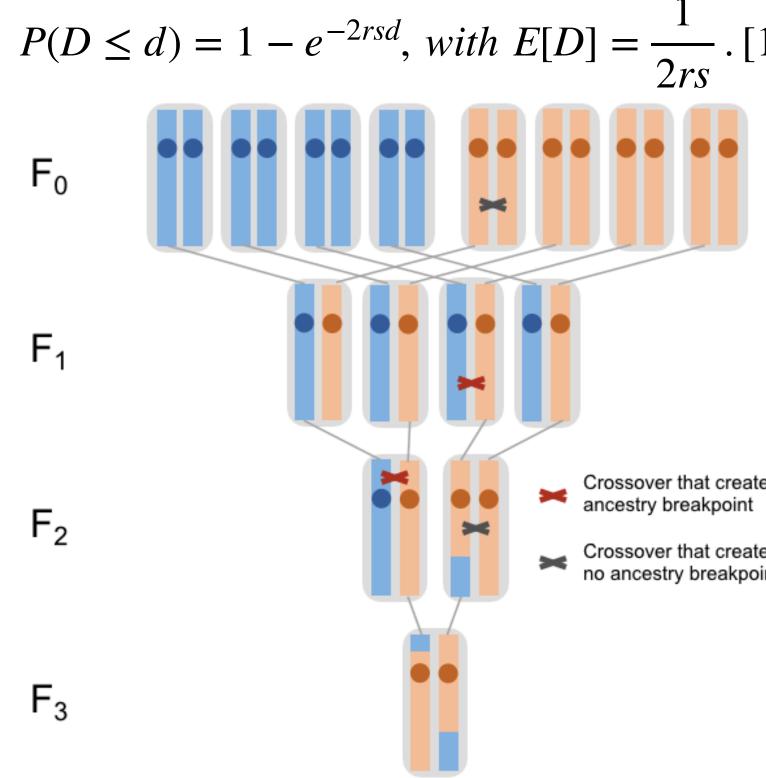
BSA is a mapping approach that combines certain ideas from x(t) = 2sx(t) = 2swould thus be: linkage mapping and GWAS. It starts from two parental strains of contrasting phenotypes. These strains are then crossed to **Solution Analysis & Numerical Validation** generate an F_1 population, which is further interbred for several Finite Population: suppose we have a diploid population of size N under the Wrightgenerations while maintaining a sufficiently large population size Fisher model, and we denote the number of haploid lineages as x(n) at generation n. We conducted forward-in-time, individual-based simulations of a BSA experiment to evaluate to allow recombination to break up linkage from the two parental Hence, in the final generation F_t with s diploid samples, we have x(t)=2s (see figure in the the accuracy of our analytical results. We modeled a trait determined by a single QTL, located strains. In the final generation, two pools of individuals are third column). at the center of a chromosome of length 100 Mbp with a uniform recombination rate of $r = 10^{-8}$ selected from the tails of the phenotypic distribution. The alleles Recursive Exact Solution: the expected number of lineages in generation n can be per bp and generation. The free parameters of our simulation model are the sample size (s), the responsible for trait differences (as well as any alleles linked to estimated based on results of occupancy distributions using a recursive expression: population size (N), and the length of the BSA experiment (t). Simulations were implemented in them) should then show characteristic frequency differences SLIM, using tree sequence recording to track ancestry segments along each chromosome (this between the two pools, while alleles at other loci should still be *et al*. 2011) allowed us to directly detect the true location of ancestry breakpoints). segregating at similar frequencies to those expected in the F_1 .



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BSA Genomic Resolution





Infinite Population vs Finite Population Model

Infinite Population: extend the processes from above to s sampled individuals from the F_t . As we neglect drift here, all lineages are independent of each other. The expect D F_t **b**

$$E[D] = \frac{1}{rst} . [2]$$

$$E[x(n-1)|x(n) = i] = N - N(1 - \frac{1}{N})^{i} . [3] (Maruvka)$$

With the above expression, we can then sum up the expected number of lineages in each generation to calculate the expected total length (7) of the genealogy, where all the ancestry breakpoints could be generated, and estimate the expected D in the finite population model as:

S:

$$E[T] = \sum_{i=3}^{l} \frac{E[x(n)]}{2} + E[x(2)] \Rightarrow E[D] = \frac{1}{rE[T]}$$

The factor of 1/2 in the above expression characterizes the probability of heterozygosity at any given genomic position in generation n>2, where ancestry breakpoints could be generated. The only exception is for the lineages in generation n=2, whose parents are all heterozygous at any genomic position from the BSA experiment setup. Approximate Closed-form Solution: by expressing the recursive solution in a differential equation, we can also solve it for a deterministic approximation of the expected number of lineages at generation *n*:

$$[x(n)] = \frac{2s}{(2s - (2s - 1)e^{-\frac{t - n}{4N}})} . [5] (Maruvka \ et \ a$$

The expected total length (T) of the genealogy in this case can then be integrated from 0 to *t*:

$$E[T] = \frac{1}{2} \int_0^t E[x(n)] \Rightarrow E[D] = \frac{1}{rE[T]} = \frac{1}{2rNln(2s(e^{\frac{t}{4N}}))}$$

Model Comparison

-. [4]

al. 2011)

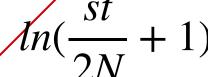
(-1) + 1)

[6]:

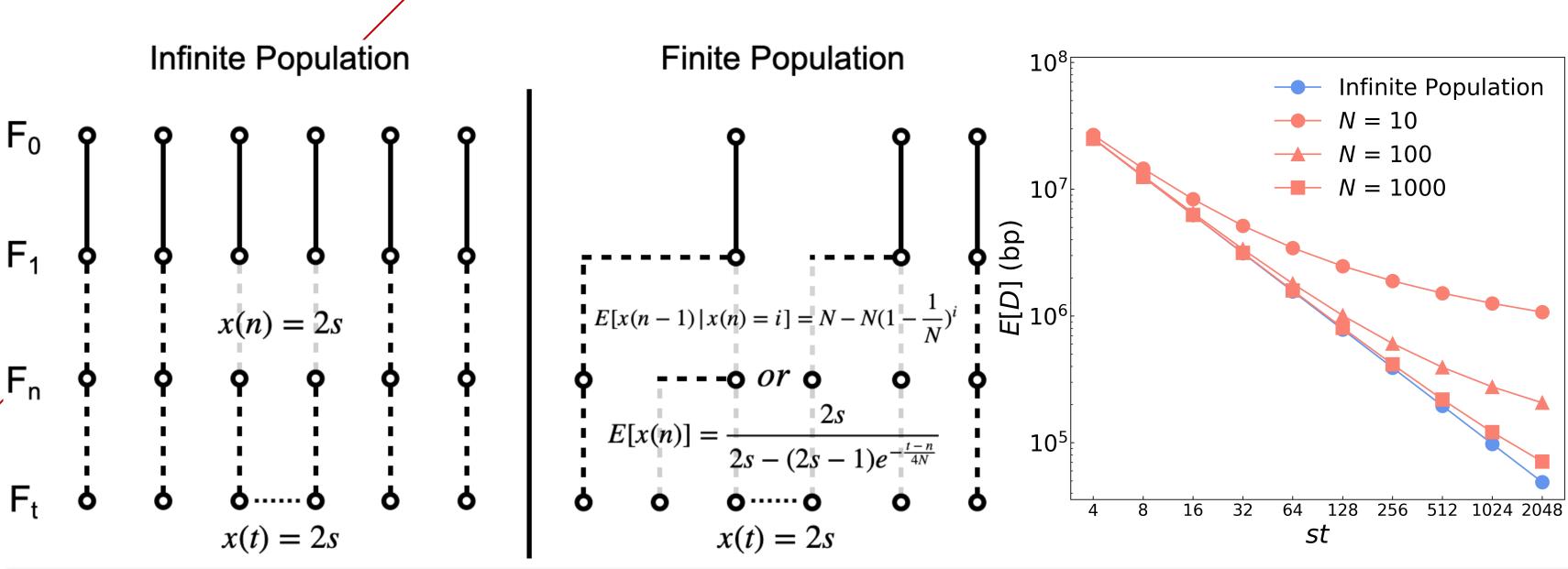
We now take a closer look at the expected mapping resolution derived in Eq. [6] and discuss how it relates to the infinite population model. When $t \ll 2N$, it specifies a regime where the probability that a given pair of lineages coalesces over the course of the experiment is still small. Under this assumption, we can perform a Taylor approximation of the exponential in Eq. $ln(2s(e^{\frac{t}{4N}} - 1) + 1) \approx ln(\frac{st}{2N} + 1) \Rightarrow E[D] \approx \frac{1}{2rMn(\frac{st}{2N} + 1)}.[7]$

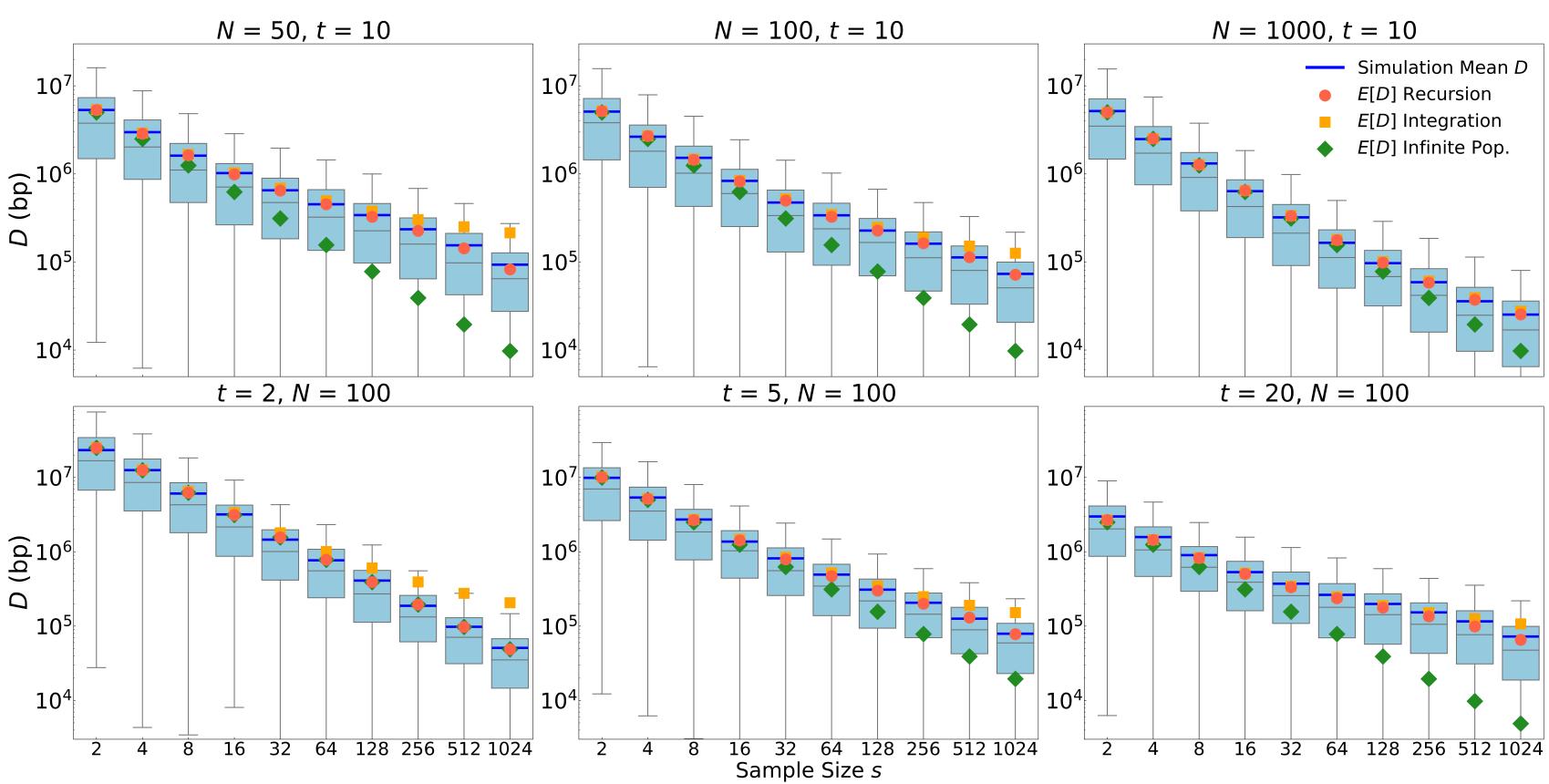
Eq. [7] shows how the infinite and finite population model, mapping resolution recombination rate, sample size, and ler mapping resolution is still inversely prop sample size and experiment length are now attenuated by a logarithm.

We further note that s and t enter Eq. [7] only in the form of the product st. Thus, varying each of these two parameters by the same factor is expected to produce a similar impact on the expected mapping resolution (as long as $t \ll 2N$ still holds). Eq. [7] also shows us where these effects start to become relevant. If st << 2N, we can further approximate:



Thus, the infinite and finite population models nicely converge in this regime.





$$ext{)} \approx rac{st}{2N} \Rightarrow E[D] \approx rac{1}{rst} . [8]$$