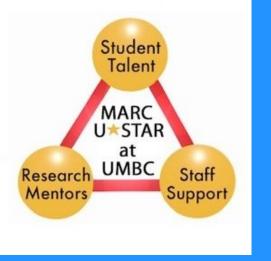


Evidence for Conserved Mechanisms of Neurulation in the Zebrafish Forebrain Dominique Brooks, Allyson Caldwell, Jonathan Werner, Rachel Brewster PhD

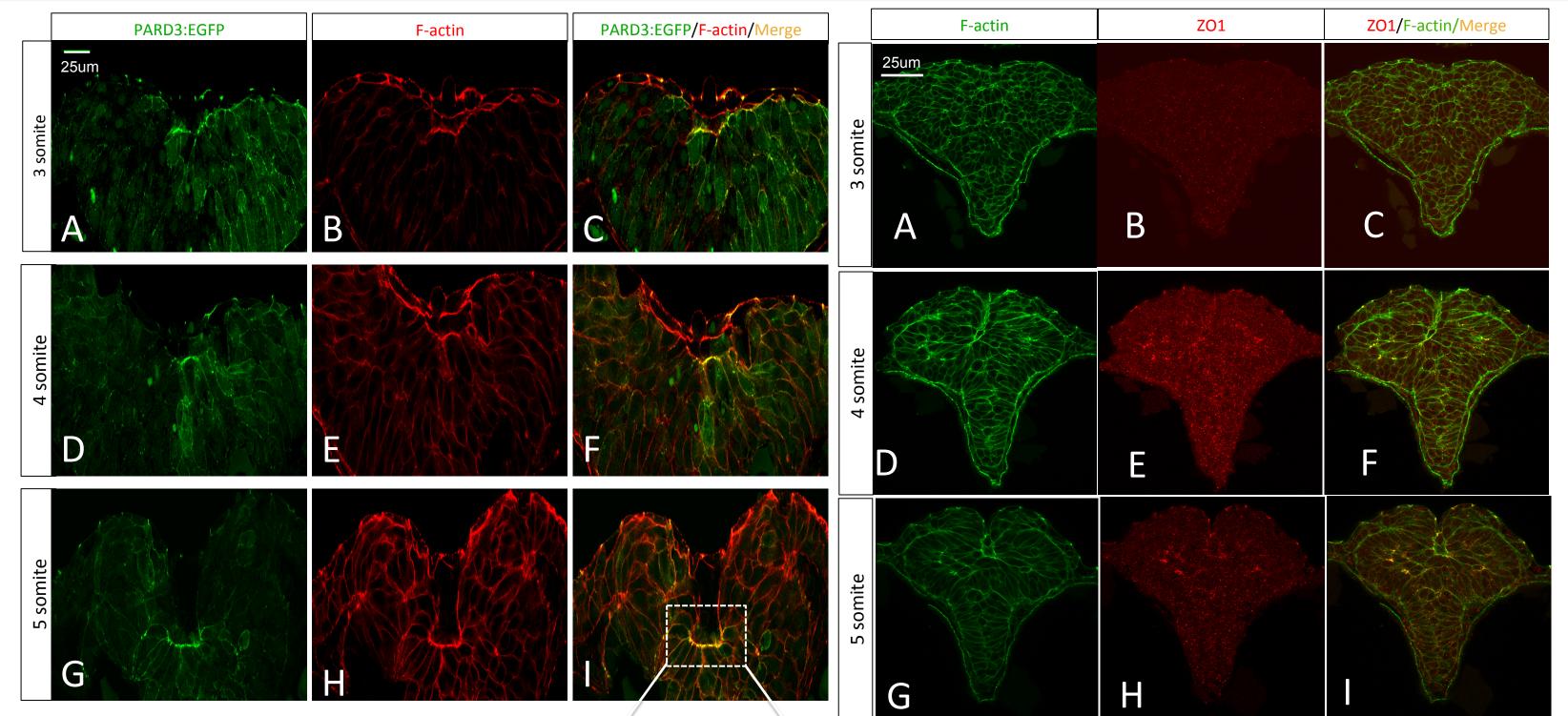


Abstract

The formation of the neural tube, the developmental precursor of the central nervous system, is facilitated by the bending and folding of the neuroepithelium, a process termed neurulation. Neural tube defects (NTD) are the most common type of birth defect in humans, primarily caused by improper neural tube formation. Investigating the cellular and molecular mechanisms that drive neurulation will help identify genetic risk factors for NTDs. The morphogenesis of the neural tube is facilitated by the formation of hingepoints, subsets of neuroepithelial cells that undergo apical constriction to form a wedge shape. Apical constriction occurs when a cell acquires a molecularly defined apical surface through apically polarized tight junction molecules, such as zona occludens (ZO1) and PARD3, in addition to the recruitment of an actomyosin contractile ring. Live imaging of hingepoint dynamics would further advance our understanding of hingepoint formation; however, it is difficult to perform in traditional model organisms. In contrast, the transparency and early accessibility of zebrafish embryos make them amenable to live imaging. Hingepoints were previously not reported in the zebrafish neuroepithelium, but we have recently found evidence for the presence of these structures in the forebrain. Using immunolabeling and confocal microscopy, we show the apical localization of ZO1 and PARD3 in a cluster of medial, wedge-shaped cells in the anterior neuroepithelium. Furthermore, we reveal that disruption of the apical actomyosin contractile ring, using the myosin inhibitor blebbistatin and myosin morpholinos, prevents apical constriction. These findings provide evidence for the presence of hingepoints in zebrafish and highlight the conservation of neural tube morphogenesis in teleosts, which pave the way for future investigations on the cellular and genetic basis of NTDs using zebrafish as a model organism.

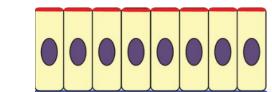
PARD3 and ZO1 are apically localized in a cluster of medial, wedge-shaped cells in the anterior neuroepithelium.

Results



Background

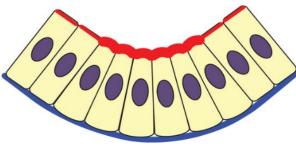
Apical Constriction and Acto-myosin Contractile Ring Facilitate Neural Plate Bending and Folding



[•] Red indicates an accumulation of polarit molecules which allows for the cell to be apically/basally polarized.



* During this transition state, red indicates an accumulation of actin/myosin allows for the convergence of the neural tissue



contractile ring is crucial to proper apical constriction, hence essential to neural plate bending and folding.

During apical

shaped.

constriction, cells

An acto-myosin

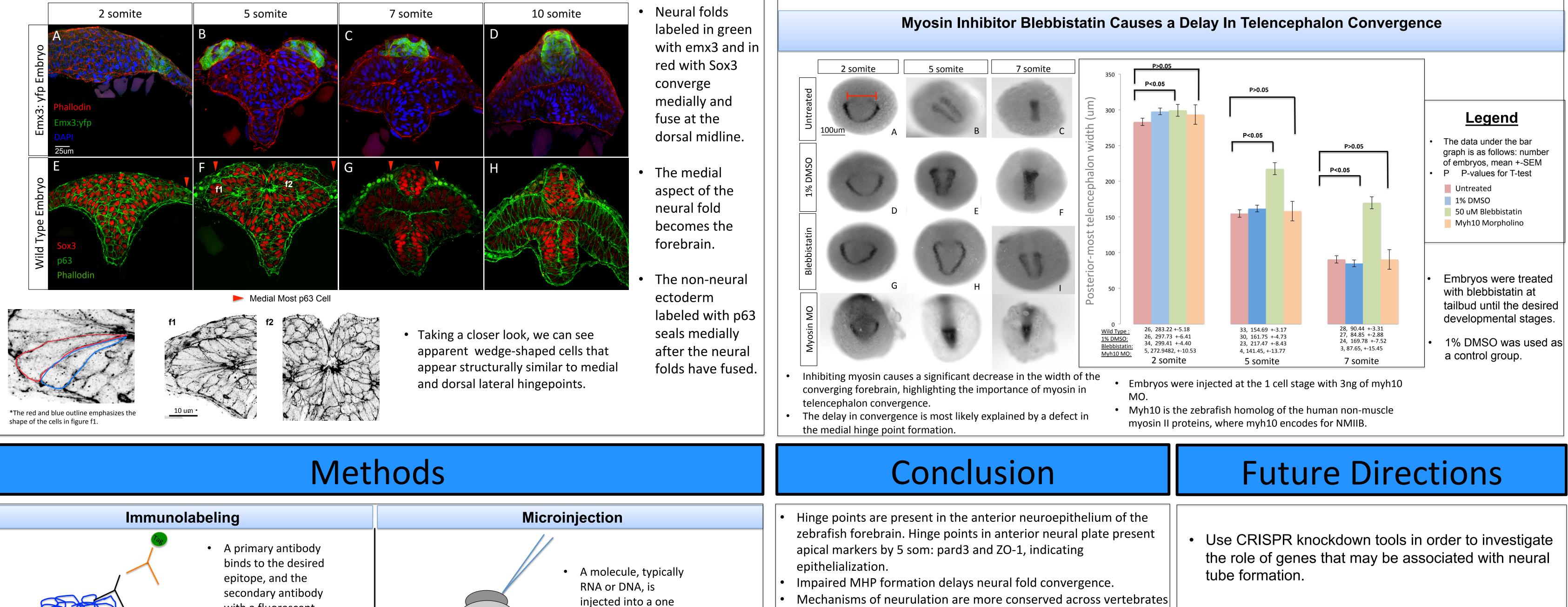
Pearl, E. J., Li, J., & Green, J. B. A. 2017 Medial and Dorsal Lateral Folds are Noticeably Present within The Zebrafish Forebrain

2-3 somite 4 somite 5 somite 6 somite 7 somite actively shape change from cuboidal to wedge-

Dorsal Ventral Fold Medial Fold

• Optical cross sections showing the developmental progression of the forebrain tissue , highlighting interesting structures in the medial and dorsal lateral regions that appear to be neural folds.

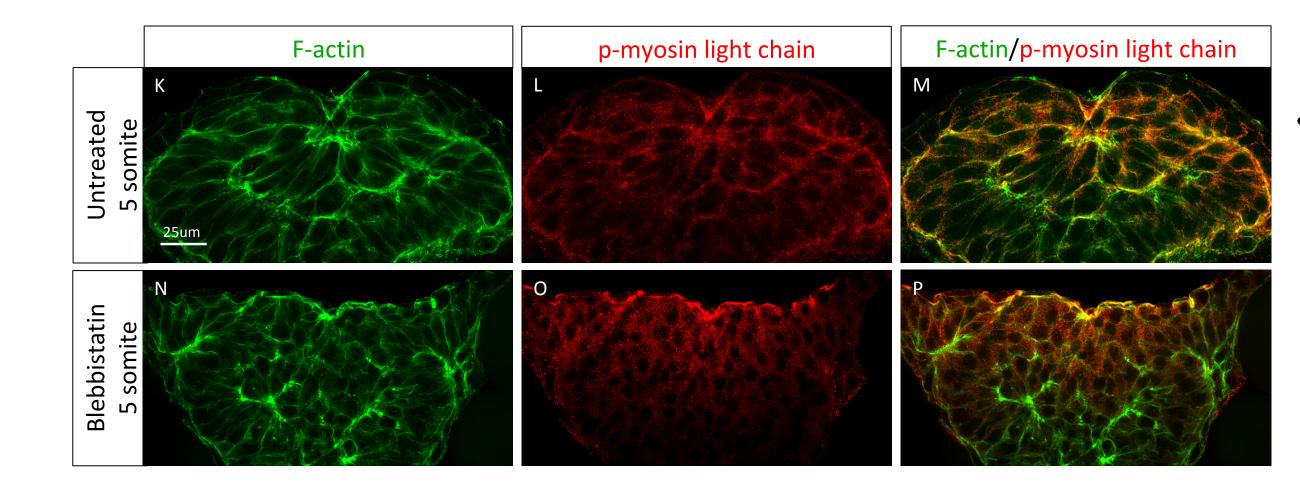
Neural Folds Migrate and Move Medially to Shape the Zebrafish Forebrain



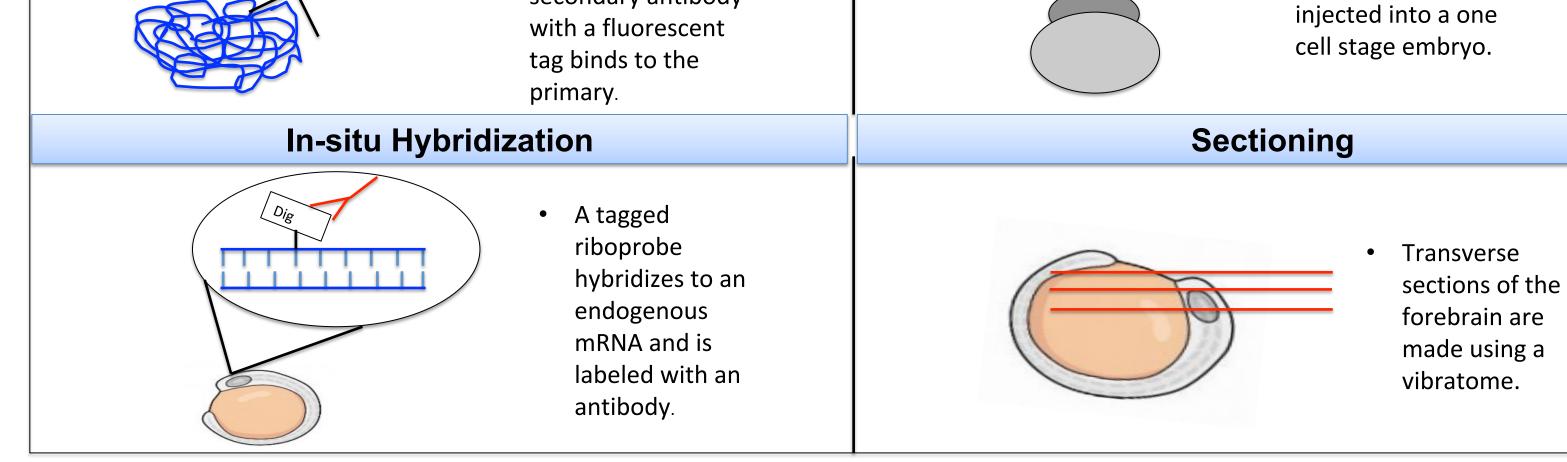
PARD3 is concentrated in a cluster of medial wedge-shaped cells, demonstrating that these cells are apically polarized.

- By 4 som, the lateral hinge points appear to be enriched with ZO-1. By 5 som, the medial deep layer also shows faint enrichment of ZO-1, indicating the cells are apically polarized.
- Establishment of apico-basal polarity is a required step preceding recruitment of contractile machinery.

Molecular characterization of wedge-shaped cells: apical localization of actin and myosin



Apical constriction appears to be facilitated by contraction of an acto-myosin contractile ring.



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